Regulation of Circulating Progenitor Cells in Left Ventricular Dysfunction

Barry A. Boilson, MB, BCh, MD; Katarina Larsen, MD; Adriana Harbuzariu, MD; Sinny Delacroix, MD; Josef Korinek, MD; Harald Froehlich, MD; Kent R. Bailey, PhD; Christopher G. Scott, MS; Brian P. Shapiro, MD; Guido Boerrigter, MD; Horng H. Chen, MD; Margaret M. Redfield, MD; John C. Burnett, Jr, MD; Robert D. Simari, MD

Background—Reductions in numbers of circulating progenitor cells (CD34+ cell subsets) have been demonstrated in patients at risk for, or in the presence of, cardiovascular disease. The mediators of these reductions remain undefined. To determine whether neurohumoral factors might regulate circulating CD34+ cell subsets in vivo, we studied complementary canine models of left ventricular (LV) dysfunction.

Methods and Results—A pacing model of severe LV dysfunction and a hypertensive renal wrap model in which dogs were randomized to receive deoxycorticosterone acetate (DOCA) were studied. Circulating CD34+ cell subsets including hematopoietic precursor cells (HPCs: CD34+/CD45dim/VEGFR2−) and endothelial progenitor cells (EPCs: CD34+/CD45−/VEGFR2+) were quantified. Additionally, the effect of mineralocorticoid excess on circulating progenitor cells in normal dogs was studied. The majority of circulating CD34+ cells expressed CD45dimly and did not express VEGFR2, consistent with an HPC phenotype. HPCs were decreased in response to pacing, and this decrease correlated with plasma aldosterone levels (Spearman rank correlation = −0.67, P=0.03). In the hypertensive renal wrap model, administration of DOCA resulted in decreased HPCs. No changes were seen in EPCs in either model. Normal dogs treated with DOCA exhibited a decrease in HPCs in peripheral blood but not bone marrow associated with decreased telomerase activity.

Conclusions—This is the first study to demonstrate that mineralocorticoid excess, either endogenous or exogenous, results in reduction in HPCs. These data suggest that mineralocorticoids may induce accelerated senescence of progenitor cells, leading to their reduced survival and decline in numbers. (Circ Heart Fail. 2010;3:635-642.)

Key Words: heart failure ▪ hypertension ▪ progenitor cells ▪ aldosterone

There is growing evidence that bone marrow-derived progenitor cells may have reparative roles in cardiac and vascular disease and may serve as diagnostic and prognostic biomarkers.1,2 Although the exact nature of these cells remains a topic of great interest, much of the work has focused on cells that are isolated by expression of CD34 or generated in vitro from circulating cells that express CD34.3 Cells originally identified in vitro as endothelial progenitor cells (EPCs: CD34+/CD45−/VEGFR2+) probably represent cells of hematopoietic origin that have assumed an endothelial phenotype including expression of VEGFR2.4,5 These cells have been demonstrated to be capable of important functional effects that are probably due to paracrine mediators.4 Circulating CD34+ hematopoietic precursor cells (HPCs: CD34+/CD45dim/VEGFR2−) are distinct from EPCs by the lack of VEGFR2 expression and by the expression of CD45 and may be the source of so-called outgrowth endothelial cells in culture.6 We recently demonstrated in humans that rigorous characterization of CD34+ cell subsets provides distinct correlates of cardiovascular risk.7 In fact, it was the HPC rather than the circulating EPC population that was downregulated in patients with coronary endothelial dysfunction. This distinction may reflect the importance of hematopoietic precursors in the early stages of atherosclerosis. Although there is growing evidence that circulating progenitor cells may be regulated in heart failure (upregulated in class 1 and downregulated in severe congestive heart failure),8 the goal of the current study was to utilize large animal models of cardiac dysfunction to assess the role of neurohumoral activation in the downregulation of circulating CD34+ cells.

Clinical Perspective on p 642

Activation of the renin-angiotensin-aldosterone system occurs early in the pathophysiology of congestive cardiac...
It has recently been shown that aldosterone impairs progenitor cells in vitro. We therefore hypothesized that mineralocorticoid excess might be associated with reductions in circulating progenitor cell counts in animal models of cardiovascular disease and that this association may be mediated by reduction in production or survival of circulating progenitor cells. Thus, we studied distinct but complementary canine models of cardiovascular disease with or without endogenous and exogenous mineralocorticoid excess. Further, to understand whether mineralocorticoid excess affected abundance or survival of progenitor cells, peripheral and bone marrow progenitor cells were studied in normal dogs before and after exogenous administration of mineralocorticoid. To quantify progenitor cells in these models, we adapted established methodology from human studies and used canine-specific reagents to study HPCs and EPCs.

Methods

Animal Models
Established canine models of cardiovascular disease were used in this study. All animal studies were performed in accordance with the Animal Welfare Act and approved by the Mayo Clinic Institutional Animal Care and Use Committee.

Pacing Model
The paced systolic dysfunction model is a well-studied model of dilated cardiomyopathy. Severe congestive cardiac failure was induced in 10 male mongrel dogs (weight, 20 to 30 kg) by rapid right ventricular pacing at 240 bpm, as described previously in detail. Blood was drawn at baseline and on the day of acute study in all animals. After 10 days of pacing, cardiorenal parameters were assessed in an acute study according to a protocol previously described. Plasma aldosterone and atrial and B-type natriuretic peptide (ANP and BNP, respectively) were measured by radioimmunoassay. It has recently been shown that aldosterone impairs progenitor cells in vitro. We therefore hypothesized that mineralocorticoid excess might be associated with reductions in circulating progenitor cell counts in animal models of cardiovascular disease and that this association may be mediated by reduction in production or survival of circulating progenitor cells. To understand whether mineralocorticoid excess affected abundance or survival of progenitor cells, peripheral and bone marrow progenitor cells were studied in normal dogs before and after exogenous administration of mineralocorticoid. To quantify progenitor cells in these models, we adapted established methodology from human studies and used canine-specific reagents to study HPCs and EPCs.

Progressive Hypertensive Heart Disease With and Without Mineralocorticoid Excess
Ten dogs ages 7 to 10 years were studied as previously described. These dogs were the last 10 dogs studied in a recently published study. In brief, all underwent a midline abdominal incision under general anesthesia with wrapping of both kidneys as previously described. All dogs were also instrumented with an indwelling aortic catheter through the femoral artery for blood pressure measurement. After development of hypertension (5 weeks after renal wrap surgery), dogs were randomized to receive deoxycorticosterone acetate (1 mg · kg⁻¹ · d⁻¹ for 3 weeks; RW DOCA) or no additional treatment (RW control). As described in this model, DOCA increased conscious blood pressure and left ventricular (LV) diastolic stiffness without a change in LV ejection fraction.

DOCA in Normal Dogs
An additional group of 6 young normal dogs underwent administration of DOCA at a similar dose of 1 mg · kg⁻¹ · d⁻¹ IM for 10 days to ascertain a mechanism for the observations. Blood and bone marrow (by aspirate from the humeral head) were analyzed at baseline and at 10 days for CD34⁺ progenitor cell count. Blood pressures were measured at 10 days at the time of acute hemodynamic study.

Flow Cytometry
Peripheral blood and bone marrow–derived cells were incubated with fluorochrome-conjugated antibodies to CD34–fluoroscein (R&D Systems), mineralocorticoid receptor (Abcam, Cambridge, Mass), VEGFR2–APC (R&D Systems), and a biotinylated rat anti-canine CD45 antibody (R&D Systems) subsequently labeled with Streptavidin–PerCP (BD Biosciences). Murine IgG₁ (R&D Systems) conjugated to Alexa 488, PE (Molecular Probes) and Rat anti-mouse PerCP (BD Biosciences) was used as isotype controls as well as IgG₁–APC from Abd Serotec. HPC (CD34⁺ CD45⁻ VEGFR2⁻) cells were enumerated using the ISHAGE single platform sequential gating strategy, which is an internationally validated standard (supplemental Figure 1). EPCs (CD34⁺ CD45⁻, and VEGFR2⁺ cells) were enumerated by sequential gating on CD34⁺, CD45⁻, and VEGFR2⁺ cells (supplemental Figure 2). Progenitor cell counts are expressed as percent total leukocyte count or absolute cell counts as indicated. Demonstration of the anti-VEGFR2 antibody on canine aortic endothelial cells was performed as described below in supplemental Figure 3.

Isolation of CD34⁺ Cells From Blood and Bone Marrow (Uninstrumented Normal Dogs)
At baseline, 20 mL of peripheral blood was drawn from the external jugular vein, and animals were anesthetized briefly with ketamine and diazepam followed by intubation and administration of an isoflurane/oxygen mixture via the endotracheal tube. The upper humeral area was shaved, prepped, and draped, and a 13-gauge bone marrow aspiration/biopsy needle (Medical Device Technologies, Fla) was advanced through the distal end of the greater tubercle of the humerus at an angle of 45° facing distally until the marrow cavity was aspirated (2 to 3 ml). After aspirates of marrow were aspirated and transferred to an EDTA tube on ice. Animals were then allowed to recover, and DOCA administration was commenced 48 hours later. An identical blood draw was performed before the acute study. After induction and intubation, bone marrow aspiration was performed again as above before commencement of invasive hemodynamic assessment. Blood was processed by density gradient centrifugation with extraction of buffy coat as previously described. Bone marrow was treated with 0.014% collagenase solution for 30 minutes, and the resulting cell suspension was diluted with PBS and layered over equal volumes of Ficoll-Paque PLUS (Amersham Biosciences, Uppsala, Sweden) for buffy coat extraction as described below.

Cells from blood or marrow were washed twice, counted, and 14-blocked with 1 µg of mouse IgG per 10⁶ cells. Approximately 7 million cells were kept for flow cytometry, which was performed as described above. The remainder (approximately 10⁸ cells) was incubated with CD34–APC antibody (R&D systems) according to the manufacturer’s instructions. Cells were washed twice and recounted and then incubated with Anti-APC Microbeads (Miltenyi Biotech, Germany), as specified by the manufacturer. Cells were again washed twice and resuspended in 50 mL PBS for magnetic cell separation. Cells were separated using an AUTOMACS machine (Miltenyi Biotech) on a positive cell selection “posseld” program, collected, and pelleted for telomerase quantification as described below.

Quantification of Telomerase Activity by Real-Time Polymerase Chain Reaction
Telomerase activity was quantified using the real-time polymerase chain reaction–based Quantitative Telomerase Detection (QTD) Kit (Allied Biotech, Ijamsville, Md) according to the manufacturer’s protocol. In brief, the CD34⁺ cells from the blood were collected, washed in PBS, and centrifuged for 30 minutes at 12 000g at 4°C. The pellet was stored at −80°C until all samples were collected. All samples were run in triplicate. Positive (enclosed with the kit) and negative controls (heat inactivated product in lysis buffer) were run in triplicate. Telomerase activity was quantified using the Chain Reaction–Based Quantitative Telomerase Detection Kit. Immediately after the initial 10-minute incubation period, 100 µL of 2 × reaction mix was added and the amplification program was started. The fluorescence intensity of the samples was then measured using an iCycler (Biorad) as specified in the manufacturer’s protocol. The value is given as fold change compared to the negative control.
buffer) were included in the analysis. A standard curve for telomerase activity was generated using provided control templates. Telomerase activity is presented in relative units.

**Statistical Analysis**

Normally distributed data are reported as mean±SEM. Nonnormally distributed data are presented as median [25th percentile, 75th percentile]. Within-group comparisons were performed using the Wilcoxon signed rank test. For between-group comparisons with repeated measures, regression analysis of posttreatment counts as the y variable, treatment group as the x variable of interest, and mean pretreatment count as a covariate was undertaken. Simple associations between cell counts and neurohormonal markers were assessed by Spearman rank correlation. The level of significance was set at P<0.05.

The authors had full access to the data and take responsibility for its integrity. All authors have read and agree to the manuscript as written.

**Results**

Rapid cardiac pacing resulted in hemodynamic evidence of advanced congestive cardiac failure as evidenced by decreased cardiac output (2.2 [1.8, 2.5] L/min) and increased systemic vascular resistance (40.4 [33.8, 51.7] WU) and pulmonary capillary wedge pressure (22.5 [14.8, 25.5] mm Hg). In dogs, the majority of circulating CD34 cells was CD45dim and VEGFR2− consistent with an HPC phenotype (95.8±1.9%). There was a significant decrease in cells was CD45dim and VEGFR2− 1.9%). There was a significant decrease in cells was CD45dim and VEGFR2− after 10 days of pacing (0.09 [0.06, 0.15]% of circulating leukocytes at baseline versus 0.05 [0.04, 0.07]% at 10 days, P=0.009 by Wilcoxon signed rank test) (Figure 1A). No significant change in EPCs (CD34+/CD45+/VEGFR2+) was detected (0.002 [0, 0.005]% of circulating leukocytes at baseline versus 0 [0, 0.004]% at 10 days, P=0.49) (Figure 1B). Similar results were seen when absolute cell counts are compared (supplemental Figure 3). There was no significant change in circulating leukocyte counts, measured as the number of total CD45+ cells per buffy coat isolate 3.4 [2.6, 4.2]×10^5 cells/100 µL buffy coat at baseline versus 3.2 [2.2, 3.7]×10^5 cells/100 µL buffy coat 10 days, P=0.57).

Neurohormonal parameters analyzed in the paced dogs at the time of acute hemodynamic study revealed a significant inverse correlation between the change in HPCs over the 10-day pacing period and the plasma aldosterone level (Spearman rank correlation=−0.67, P=0.03) (Table 1 and supplemental Figure 4). No significant correlation was noted between changes in HPCs and serum ANP and BNP levels, nor did changes in EPCs correlate with plasma aldosterone, ANP, or BNP levels. These data suggest that elevation in serum aldosterone levels is associated, at least in part, with changes in circulating cell subsets.

On the basis of the association between change in cell counts and aldosterone levels, we chose to further study the relationship between mineralocorticoid excess and CD34+ cell subsets in a model of hypertension that uses bilateral RW and administration of DOCA. DOCA has been shown to have important effects on LV and vascular form and function in this model.17 The hemodynamic profiles of RW control and RW DOCA groups at the time of acute study are shown in Table 2. Again, the majority of circulating CD34+ cells was CD45dim and VEGFR2−, consistent with an HPC phenotype (94.1±1.1%). When the effects of DOCA administration were analyzed at 6 and 8 weeks, regression analysis adjusting for pretreatment mean as covariate revealed the differences in cell counts between groups at 8 weeks to be statistically significant (P=0.01) (Figure 2).

To determine whether mineralocorticoid excess is sufficient to account for decreases in HPCs, uninstrumented dogs were treated with DOCA for 10 days. No significant effect on total leukocyte counts was noted after DOCA treatment (Figure 3A).

**Table 1.** Spearman Rank Correlations Between Change in Progenitor Cell Counts From Paced Dogs Over 10 Days Compared With Plasma Aldosterone Level, Canine BNP Levels, and Canine ANP Levels at 10 Days

<table>
<thead>
<tr>
<th>Change in HPC Count as % of Total Leukocytes</th>
<th>Change in EPC Count as % of Total Leukocytes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spearman Rank Correlation</td>
<td>P Value</td>
</tr>
<tr>
<td>Plasma aldosterone level, ng/mL</td>
<td>−0.67</td>
</tr>
<tr>
<td>Plasma canine BNP level, pg/mL</td>
<td>0.10</td>
</tr>
<tr>
<td>Plasma ANP level, pg/mL</td>
<td>0.15</td>
</tr>
</tbody>
</table>
However, a significant fall in HPCs as a percentage of total leukocytes was noted in peripheral blood comparing baseline values with those at 10 days (0.10 [0.07, 0.17]% to 0.02 [0.02, 0.05], \( P = 0.005 \) by Wilcoxon signed rank test) (Figure 3B) but not in bone marrow (0.11 [0.06, 0.18]% to 0.18 [0.09, 0.33]% , \( P = 0.30 \)) (Figure 3C). EPCs did not change significantly in peripheral blood or in bone marrow after treatment with DOCA for 10 days (Figure 3B and 3C, right panels).

To determine whether cells in canine buffy coat express the mineralocorticoid receptor (MR), we demonstrated mRNA for MR present in these samples (Figure 4). Furthermore, we performed FACS on canine blood and demonstrated coexpression of CD34 and MR. Immunostaining of canine bone marrow also showed coexpression of CD34 and MR.

Finally, circulating CD34+ cells demonstrated evidence of senescence with reduced telomerase activity after administration of DOCA (Figure 5). No differences in telomerase activity were seen in bone marrow--derived CD34+ cells, suggesting a peripheral effect. We were not able to perform these studies on CD34+ subsets. However, because the overwhelming majority of CD34+ cells are HPCs, we assume these findings represent this population. Taken together, DOCA may induce a decrease in circulating CD34+ cells through induction of senescence.

<table>
<thead>
<tr>
<th>RW, Control (n = 5)</th>
<th>RW, DOCA (n = 5)</th>
<th>( P ) Value‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>MAP,* mm Hg</td>
<td>138.6</td>
<td>165.2</td>
</tr>
<tr>
<td>PCWP,† mm Hg</td>
<td>9.5</td>
<td>8.9</td>
</tr>
<tr>
<td>CO,† L/min</td>
<td>2.0</td>
<td>1.8</td>
</tr>
<tr>
<td>SVR, Woods units†</td>
<td>66.0</td>
<td>40.4</td>
</tr>
</tbody>
</table>

Median values shown.
*Conscious state; †under general anesthesia; ‡Wilcoxon rank sum test.

Table 2. Hemodynamics at Acute Hemodynamic Study in RW Groups

Figure 2. Effects of RW and DOCA on progenitor cell counts. HPC and EPC counts over time with regression analysis for the effect of DOCA therapy with the mean of pretreatment values as covariate. HPC (A) and EPC (B) counts as percents of total leukocytes.
Discussion

It is now well established that progenitor cells of bone marrow origin can be detected in the circulation, and altered numbers and function of these cells have been noted in cardiovascular disease, including heart failure. However, the mechanisms involved and the implications of these findings have remained elusive. In the present study, 2 canine models of LV dysfunction were used. Specific canine reagents and analyses were used to quantify 2 populations of circulating CD34^+ cells, EPCs, and HPCs. HPC counts were decreased in a paced model of cardiac failure. No changes in EPCs were noted. Interestingly, this decrease in HPCs correlated with increases in plasma aldosterone levels. On the basis of this original finding, we hypothesized that mineralocorticoid excess may be associated with decreased circulating HPCs. In this setting, aldosterone levels may be a surrogate for increased systemic oxidative stress. When DOCA, an MR agonist, was administered to the RW model, there was a significant reduction in circulating HPC counts compared with controls. Again, no changes were seen in EPC counts. DOCA was also administered to normal dogs, resulting in a similar reduction on HPCs observed after 10 days of treatment. In the latter animals, we also analyzed bone marrow before and after treatment. It was interesting to note that the reduction in HPC number seen in the
circulation was not seen in the bone marrow, suggesting a peripheral effect on cell survival.

In the present study, the majority of CD34+ cells expressed CD45dimly and did not express VEGFR2, consistent with the HPC phenotype. These HPCs were regulated by mineralocorticoid excess, whereas no effects were seen in EPCs (CD34+/CD45−/VEGFR2+ cells). The lack of effect on EPCs may be due to their relative scarcity in the population, but their levels are consistent with our study in humans.7 It should be noted that the study was limited by a lack of canine specific reagents for other hematopoietic stem cell markers such as CD133, c-kit, or sca-1. Taken together, these data suggest that CD34+ cells are regulated in models of LV dysfunction associated with mineralocorticoid excess. However, it was the HPC population that accounted for these differences. Because these HPCs may be the source of multiple relevant cell types including cells capable of assuming an endothelial phenotype, the findings have particular relevance to cardiovascular disease.

Our finding that CD34+ cells express the mineralocorticoid receptor is consistent with accumulating evidence that circulating progenitor cells express MRs28,29 and also that their growth in vitro is impaired by addition of aldosterone to the culture medium.14 These findings are made all the more compelling by the observed reversal of these effects in vitro with the addition of spironolactone. These observations underscore the possibility that aldosterone exerts specific effects on circulating progenitor cells. However, the presence of the MR alone does not prove a direct effect. The expression of 11β-hydroxysteroid dehydrogenase is also a neces-

Figure 4. Detection of MR expression. A, Reverse transcription–polymerase chain reaction for MR in peripheral blood buffy coat from normal dogs. B, FACS of canine peripheral blood for CD34 and MR (R1=CD34+) (R2=CD34+/MR+). C, Colocalization of CD34 (red, cytoplasmic staining) and MR (green, nuclear staining) in canine bone marrow. Arrow indicates cell expressing CD34 and MR.
underlying mechanisms may include effects on telomerase activity in circulating progenitor cells leading to possible telomere length shortening and accelerated cell death. The implications of these findings suggest a novel paradigm in which to consider aldosterone antagonism in human disease.

Acknowledgments

We thank Jim Tarara and the staff at the Mayo Clinic Flow Cytometric Core Facility for their assistance with sample processing for flow cytometric analysis. We also thank Gail J. Harty, Donna M. Meyer, Denise Heublein, Sharon Sandberg, Cheryl Mueske, Lauren Kleppe, and Tyra Witt for their technical assistance and to Megan E. Crouch for her assistance with preparation of the manuscript.

Sources of Funding

This study was funded by the National Institutes of Health (HL76611).

Disclosures

None.

References


Regulation of Circulating Progenitor Cells in Left Ventricular Dysfunction
Barry A. Boilson, Katarina Larsen, Adriana Harbuzariu, Sinny Delacroix, Josef Korinek, Harald Froehlich, Kent R. Bailey, Christopher G. Scott, Brian P. Shapiro, Guido Boerrigter, Horng H. Chen, Margaret M. Redfield, John C. Burnett, Jr and Robert D. Simari

*Circ Heart Fail.* 2010;3:635-642; originally published online June 23, 2010; doi: 10.1161/CIRCHEARTFAILURE.109.879437
*Circulation: Heart Failure* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2010 American Heart Association, Inc. All rights reserved.
Print ISSN: 1941-3289. Online ISSN: 1941-3297

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circheartfailure.ahajournals.org/content/3/5/635

Data Supplement (unedited) at:
http://circheartfailure.ahajournals.org/content/suppl/2010/06/23/CIRCHEARTFAILURE.109.879437.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation: Heart Failure* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the *Permissions and Rights Question and Answer* document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to *Circulation: Heart Failure* is online at:
http://circheartfailure.ahajournals.org//subscriptions/
Supplemental Material
Supplemental Figure Legends:

Supplemental Figure 1: ISHAGE enumeration of canine HPCs (CD34+ CD45\textsubscript{dim} VEGFR2- progenitor cells)

A. After incubation of cells with fluorophore-conjugated antibodies to CD45 and CD34, data are displayed as a dotplot of side scatter (SSC) vs. CD45 fluorescence. All CD45 positive events are gated in a region R1.

B. Data in R1 is displayed as a dotplot of side scatter vs. CD34 fluorescence. A CD34 positive cluster of low side scatter should be apparent which is gated R2.

C. Data from R2 are displayed in a dotplot SSC vs CD45 fluorescence. The CD45-dim population is gated R3.

D. Returning to the SSC vs CD45 dotplot and switching off all gates again, a new gate R4 is created around the homogenous, CD45 positive and low side scatter lymphocyte population.

E. R4 is displayed as a dotplot of side scatter vs. forward scatter. This shows the phenotypic characteristics of the lymphocytes and blasts, which are similar to those of hematopoetic precursors. A region R5 is created to set the boundaries of these defining phenotypic features.

F. Contents of R1 x R2 x R3 are displayed as in E. All events outside R5 are excluded from the final count, which represents the number of CD34+ CD45\textsubscript{dim} positive cells per 60,000 events. These cells are uniformly VEGFR2 negative.
Supplemental Figure 2: Enumeration of canine CD34+ CD45- VEGFR2+ progenitor cells (EPCs)

A. After incubation of cells with fluorophore-conjugated antibodies to CD45 and CD34, data are displayed as a dotplot of side scatter (SSC) vs. CD45 fluorescence. All CD45 negative events are gated in a region R1.

B. Data in R1 is displayed as a dotplot of side scatter vs. CD34 fluorescence. A CD34 positive cluster of low side scatter should be apparent which is gated R2.

C. Data from R2 are displayed in a dotplot SSC vs CD45 fluorescence. The CD34+ CD45-negative population is visible by gating on R1 * R2.

D. In a dotplot of SSC vs VEGFR2 a new gate R3 is created around all VEGFR2+ events.

E. In the same dotplot, CD34+ CD45- VEGFR2+ events are demonstrated by gating on R1 * R2 * R3. Contents of R1 x R2 x R3 are displayed in a dotplot of SSC vs. FSC (forward scatter). The low side scatter, low forward scatter nature of these cells is apparent.

Supplemental Figure 3: A: Hematopoietic (HPC) and B: endothelial progenitor cell (EPC) counts in a canine pacing model of LV dysfunction, expressed as absolute counts. Cells were enumerated from paced dogs at baseline and at 10 days, compared using a Wilcoxon signed-rank test.

Supplemental Figure 4: Neurohumoral correlates of progenitor cell counts: Spearman Rank correlation (\( \rho \)) of change in progenitor cell counts from paced dogs over 10 days compared with plasma aldosterone level (A), canine B-type natriuretic peptide (cBNP)
(B) levels, and canine atrial natriuretic peptide levels (ANP) (C) at 10 days. For clarity, correlations are shown in a linear fashion.

**Supplemental Figure 5. Immunodetection of VEGFR2 expression in canine cells.**

A. Canine aortic endothelial cells (Cell applications, Inc.) stained with control IgG. Hoechst counterstain depicts nuclei. B. Canine aortic endothelial cells stained with anti-VEGFR2 and Hoechst. C. FACS analysis of canine endothelial cells using IgG and the anti-VEGFR2 antibodies
Supplemental Figure 1

A: No gate

B: R1

C: R1 + R2

D: R1 + R2 + R3

E: R1

F: R1 + R2 + R3
Supplemental Figure 2

A

No gate

R1

B

No gate

R1 + R2

C

R1 + R2

D

No gate

R3

E

R1 + R2 + R3

F

R1 + R2 + R3
Supplemental Figure 3

A

HPC cell count/10 μL buffy coat

Baseline

Post pacing

p = 0.005

n = 10

B

EPC cell count/10 μL buffy coat

Baseline

Post pacing

p = 0.29

n = 10
Supplemental Figure 4

A

ρ = -0.67
p = 0.03

B

ρ = -0.10
p = 0.78

C

ρ = 0.15
p = 0.70

ρ = -0.19
p = 0.61
Supplemental Figure 5