Expression of the Irisin Precursor FNDC5 in Skeletal Muscle Correlates With Aerobic Exercise Performance in Patients With Heart Failure

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Background.—Exercise-induced increase in peroxisome proliferator-activated receptor-γ coactivator-1α (PGC-1α) expression has been shown to increase the expression of the fibronectin type III domain containing 5 (FNDC5) gene and thereby its product, irisin, in mice. Given that exercise intolerance is a hallmark characteristic of heart failure (HF), and because PGC-1α and irisin promote exercise benefits in animals, we hypothesized that expression of these genes relates to aerobic performance in patients with HF.

Methods and Results.—Systolic HF (left ventricular ejection fraction ≤40%) patients underwent cardiopulmonary exercise testing to evaluate aerobic performance. High versus low aerobic performance was assessed using oxygen consumption (peak V̇O₂ >14 versus ≤14 mL O₂·kg⁻¹·min⁻¹) and ventilatory efficiency (VE/V̇CO₂ slope [<34 versus ≥34]). Muscle biopsies of the vastus lateralis and real-time polymerase chain reaction were used to quantify muscle gene expression. Twenty-four patients were studied. FNDC5 (5.7±3.5 versus 3.1±1.2, P<0.05) and PGC-1α (9.9±5.9 versus 4.5±1.9, P<0.01) gene expressions were greater in the high-peak V̇O₂ group; correlation between FNDC5 and PGC-1α was significant (r=0.56, P<0.05) only in the high-peak V̇O₂ group. Similarly, FNDC5 and PGC-1α gene expression was greater in the high-performance group based on lower VE/V̇CO₂ slopes (5.8±3.6 versus 3.3±1.4, P<0.05 and 9.7±6 versus 5.3±3.4, P<0.05); FNDC5 also correlated with PGC-1α (r=0.55, P<0.05) only in the low VE/V̇CO₂ slope group.

Conclusions.—This is the first study to show that FNDC5 expression relates to functional capacity in a human HF population. Lower FNDC5 expression may underlie reduced aerobic performance in HF patients. (Circ Heart Fail. 2012;5:812–818.)

Key Words: heart failure ■ genes ■ physiology ■ biopsy
As such, this newly identified hormone has the potential to become a therapeutic target that may improve functional performance that is often limited in HF patients.\(^2,3\)

Although the initial description of FNDC5 expression and irisin production by Boström et al\(^9\) was made in mice, there is reason to believe that irisin also affects human physiology because the amino acid sequence of the molecule is identical between the species. It is theorized that FNDC5 may have similar roles in humans as in mice, but it is still not known how FNDC5 expression is controlled in human populations, or how it varies in different physiologic or pathophysiologic conditions.

In this analysis, we assessed PGC1-α and FNDC5 gene expression in relation to peak VO\(_2\) and the VE/V\(_{\text{CO}_2}\) slope,\(^17\) two well-established cardiopulmonary exercise test (CPX) measures of aerobic exercise performance. α and FNDC5 gene expression in relation to peak VO\(_2\) and the (VE/V\(_{\text{CO}_2}\)) slope, two well-established cardiopulmonary exercise test (CPX) measures of aerobic exercise performance. This is a potentially important area of research particularly because of the notorious decrements of functional capacity seen among HF patients in association with skeletal muscle abnormalities. Reduced irisin may be a significant component of HF-related skeletal muscle changes that influence functional decline.

**Methods**

Clinically stable male systolic (left ventricular ejection fraction ≤40%) HF patients aged ≥20 years were enrolled. All were on a standard regimen of evidence-based HF therapy, including a β-blocker and angiotensin converting enzyme-inhibitor or angiotensin receptor blocker and were euvolemic at the time of assessment. In addition, each patient had an echocardiogram within 6 months of enrollment to confirm their left ventricular ejection fractions were ≤40% at the time of the study assessments. Mitral valve regurgitation was assessed based on trace, mild, moderate, or severe regurgitation (categorized as 0, 1, 2, or 3, respectively) and is reported as an average. Patients were excluded if they had severe valvular disease, neurological disorders, pulmonary disease, or musculoskeletal problems, which could have confounded functional assessments. This study was approved by the local Veterans Affairs Institutional Review Board and all subjects signed an informed consent before participation.

**Aerobic Exercise Testing**

All patients performed a symptom-limited CPX test on a motorized treadmill (Bari-mill Woodway, Waukesha, WI) using a modified Balke protocol.\(^18\) The ventilatory expired gas analysis system (MedGraphics BreezeSuite St. Paul, MN) was calibrated according to manufacturer’s specifications before each test. Continuous gas exchange, telemetry, blood pressure, rating of perceived exertion, and oxygen saturation were assessed for each patient 1 minute before, during, and 4 minutes after the exercise test in standard clinical fashion. Both peak VO\(_2\) and the VE/V\(_{\text{CO}_2}\) slope were used to assess aerobic performance. Just as peak VO\(_2\) manifests both central (cardiac and pulmonary) and peripheral (skeletal muscle and peripheral vascular) physiologic processes,\(^19-22\) the VE/V\(_{\text{CO}_2}\) slope reflects a broad array of physiologic systems, including skeletal muscle chemo- and ergoreflexes and cardiopulmonary coupling.\(^21-24\)

Aerobic indices including peak VO\(_2\) (quantified as the highest 30-second averaged value during the last stage of exercise) and the VE/V\(_{\text{CO}_2}\) slope were evaluated. VE and V\(_{\text{CO}_2}\) values, acquired from the initiation of exercise to peak exercise (as 30-second averages), were entered into a spreadsheet software (Microsoft Excel, Microsoft Corp., Bellevue, WA) to calculate the VE/V\(_{\text{CO}_2}\) slope via least squares linear regression (y=m\(x\) + b, m=slope). All subjects achieved a respiratory exchange ratio of >1.0 at peak exercise, an indication of good exercise effort.\(^5\)

**Skeletal Muscle Biopsy**

Within 1 week after the CPX test, a skeletal muscle biopsy of the vastus lateralis muscle in the left leg was performed using a 5-mm Bergstrom muscle biopsy trochar. Biopsy samples were stored in RNALater (Qiagen, Valencia, CA) at −80°C until gene-expression analyses were conducted.

**Gene-Expression Analyses**

Total skeletal muscle mRNA was extracted from muscle biopsies, using Trizol reagent according to the manufacturer’s instructions. Total RNA was converted to cDNA by reverse transcription using SuperScript reverse transcriptase (Stratagene, Carlsbad, CA) and oligo dT primers in standard reactions according to the manufacturer’s recommendations. Gene-expression levels were determined from diluted cDNA samples (1:100) by real-time polymerase chain reaction using the StepOnePlus analyzer (Applied Biosystems, Foster City, CA). Multiplexed amplification reactions were performed using glyceraldehyde-3-phosphate-dehydrogenase as an endogenous control (glyceraldehyde-3-phosphate-dehydrogenase primers/VIC-labeled probe Applied Biosystems # 4310884E) using the TaqMan Universal PCR Master Mix reagents Kit (#4304437, Applied Biosystems). The following amplification settings were used: stage 1 (denaturation): 95°C for 10 minutes and stage 2 (polymerase chain reaction): 95°C for 15 seconds and 60°C for 60 seconds for 40 cycles. The threshold cycle values for each reaction were transferred to a Microsoft Excel spreadsheet and calculation of relative gene expression (to glyceraldehyde-3-phosphate-dehydrogenase) was performed from this data according to published algorithms (TaqMan Cytokine Gene Expression Plate 1 protocol, Applied Biosystems).\(^32\) For each gene studied, all biopsy samples were analyzed on a single 96-well plate. Expression of the following genes were analyzed using FAM-labeled probe/primer sets purchased from Applied Biosystems: PGC-1α (Hs01016724_m1) and FNDC5 (Hs00401006_m1).

**Statistical Analysis**

Statistical Analysis System (SAS) software version 9.0 (SAS, Cary, NC) was used to analyze the data; values are reported as means±SD unless otherwise indicated. The overall group was dichotomized into high versus low aerobic capacity according to clinically established thresholds for peak VO\(_2\) (>14 versus ≤14 mL O\(_2\)·kg\(^{-1}\)·min\(^{-1}\))\(^19\) and the VE/V\(_{\text{CO}_2}\) slope (<34 versus ≥34).\(^26\) Comparisons of high (>14 mL O\(_2\)·kg\(^{-1}\)·min\(^{-1}\)) or VE/V\(_{\text{CO}_2}\) slope (<34 versus low ≤14 mL O\(_2\)·kg\(^{-1}\)·min\(^{-1}\)) or VE/V\(_{\text{CO}_2}\) slope ≥34) performance among HF patients was determined using unpaired t tests, whereas evaluations of the correlation coefficients in the overall group as well as subgroups according to CPX performance were conducted by Pearson correlations. \(P<0.05\) was used to define statistical significance for all tests.

**Results**

Twenty-four male systolic HF patients Twenty-four male systolic HF patients (mean age 67.2±9.2 years, age range 50–86) were evaluated. Mean left ventricular ejection fraction was 29.5±7.7%, range 15% to 40%.

Table 2 lists echocardiography and CPX aerobic indices. Performance stratifications were based on peak VO\(_2\) (>14 versus ≤14 mL O\(_2\)·kg\(^{-1}\)·min\(^{-1}\) [n=15 and 9, respectively] and VE/V\(_{\text{CO}_2}\) <34 versus ≥34 [n=14 and 10, respectively]).\(^19,26\)

Table 2 lists echocardiography and CPX aerobic indices. In general, patients had depressed systolic function by echocardiography but no differences between groups with respect to morphology or left ventricular function. Although pulmonary pressure (pulmonary artery systolic pressure [PASP]) was similar between the high- versus low-peak VO\(_2\) groups,
Table 1.  Key Demographic Characteristics

<table>
<thead>
<tr>
<th></th>
<th>Entire Cohort (n=24)</th>
<th>Peak Vo₂ (mL O₂·kg⁻¹·min⁻¹)</th>
<th>VE/Vco₂ Slope</th>
<th>P value</th>
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<tbody>
<tr>
<td></td>
<td>(n=15)</td>
<td>(n=9)</td>
<td>P value</td>
<td>(n=14)</td>
</tr>
<tr>
<td>Age, yr</td>
<td>67.2±9.3</td>
<td>64.7±7.1</td>
<td>71.4±11.4</td>
<td>NS</td>
</tr>
<tr>
<td>Height, m</td>
<td>1.7±0.07</td>
<td>1.7±0.05</td>
<td>1.7±0.1</td>
<td>NS</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>88.3±29.3</td>
<td>87.7±19.3</td>
<td>89.4±42.5</td>
<td>NS</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>29.3±7.8</td>
<td>29.2±5.1</td>
<td>29.6±11.3</td>
<td>NS</td>
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<td>NT-proBNP</td>
<td>1518.3±18010.5</td>
<td>894.1±693</td>
<td>2558.7±2575.9</td>
<td>NS</td>
</tr>
<tr>
<td>Medications (n; % within each group)</td>
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<td></td>
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<tr>
<td>β-blocker</td>
<td>18 (75)</td>
<td>11 (73)</td>
<td>7 (78)</td>
<td>NS</td>
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<tr>
<td>ACE inhibitor</td>
<td>19 (79.2)</td>
<td>13 (87)</td>
<td>6 (67)</td>
<td>NS</td>
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<tr>
<td>Statin</td>
<td>20 (83.3)</td>
<td>13 (87)</td>
<td>7 (78)</td>
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<tr>
<td>ARB</td>
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<td>1 (7)</td>
<td>1 (11)</td>
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<td>Diuretic</td>
<td>17 (70.8)</td>
<td>9 (60)</td>
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<td>NYHA classification</td>
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<td>I</td>
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<td>4</td>
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<td>NS</td>
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Table 2.  Hemodynamic and Cardiopulmonary Indices

<table>
<thead>
<tr>
<th></th>
<th>Entire Cohort (n=24)</th>
<th>Peak Vo₂ (mL O₂·kg⁻¹·min⁻¹)</th>
<th>VE/Vco₂ Slope</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(n=15)</td>
<td>(n=9)</td>
<td>P value</td>
<td>(n=14)</td>
</tr>
<tr>
<td>Echocardiography parameters</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>LVEF, %</td>
<td>29.5±7.7</td>
<td>28.6±8.4</td>
<td>31.3±6.4</td>
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</tr>
<tr>
<td>LVIdd, cm</td>
<td>5.8±0.8</td>
<td>5.9±0.9</td>
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<td>NS</td>
</tr>
<tr>
<td>FS</td>
<td>15.1±7.6</td>
<td>16±7.6</td>
<td>13.6±6.3</td>
<td>NS</td>
</tr>
<tr>
<td>VpAP, mmHg</td>
<td>26.5±7.7</td>
<td>25.7±8</td>
<td>27.7±7.8</td>
<td>NS</td>
</tr>
<tr>
<td>MV regurgitation</td>
<td>0.3±0.7</td>
<td>0.2±0.6</td>
<td>0.6±0.9</td>
<td>NS</td>
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<tr>
<td>Cardiopulmonary indices</td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Peak Vo₂, mL O₂·kg⁻¹·min⁻¹</td>
<td>14.9±3.3</td>
<td>17.0±2.0</td>
<td>11.4±1.6</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>VE/Vco₂ slope</td>
<td>34.4±9.2</td>
<td>30.1±3.8</td>
<td>41.5±11.3</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>Peak RER</td>
<td>1.1±0.07</td>
<td>1.1±0.1</td>
<td>1.1±0.1</td>
<td>NS</td>
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</tbody>
</table>

PASP level was significantly elevated in the low aerobic performance group assessed as VE/Vco₂ slope ≥34. When defining aerobic performance based on peak Vo₂, the high-performance group was characterized by a peak Vo₂ of 17.0±3.0 versus 11.4±1.6 mL O₂·kg⁻¹·min⁻¹ in the low-performance group, P<0.0001 (Table 2). When dichotomized into 17.0±3.0 versus 11.4±1.6 mL O₂·kg⁻¹·min⁻¹ and categorized as high and low performance according to the VE/Vco₂ slope, an increase in PASP was observed in the low performance cohort assessed as VE/Vco₂ slope ≥34, or by VE/Vco₂ slope <34, had significantly greater expression of both PGC-1α and FNDC5. Within each of these 2 classifications of high-exercise performance, increased expression of FNDC5 correlated with greater expression of PGC-1α (peak Vo₂ >14 mL O₂·kg⁻¹·min⁻¹, r=0.56, P<0.05; VE/Vco₂ slope <34, r=0.55, P<0.05). Similar correlations between PGC-1α and FNDC5 were not present in the lower functional groups (peak Vo₂ <14 mL O₂·kg⁻¹·min⁻¹, r=-0.22, P=0.58; VE/Vco₂ slope >34, r=0.47, P=0.17). As demonstrated in Figure 1, FNDC5 expression correlated with PGC-1α expression in relation to the entire cohort (r=0.61 P=0.001). Correlations between PGC-1α and FNDC5 and peak Vo₂ and the VE/Vco₂ slope are illustrated in Figure 2. Although all correlations trended in the expected direction, only the relationship between PGC-1α and peak Vo₂ reached statistical significance.

Table 3 shows gene expression in relation to aerobic performance, and demonstrates significant differences between the two functional groups. shows gene expression in relation to aerobic performance, and demonstrates significant differences between the 2 functional groups. Subjects with higher aerobic performance indices, defined by either peak Vo₂ >14 mL O₂·kg⁻¹·min⁻¹, or by VE/Vco₂ slope <34, had significantly greater expression of both PGC-1α and FNDC5. Within each of these 2 classifications of high-exercise performance, increased expression of FNDC5 correlated with greater expression of PGC-1α (peak Vo₂ >14 mL O₂·kg⁻¹·min⁻¹, r=0.56, P<0.05; VE/Vco₂ slope <34, r=0.55, P<0.05). Similar correlations between PGC-1α and FNDC5 were not present in the lower functional groups (peak Vo₂ <14 mL O₂·kg⁻¹·min⁻¹, r=-0.22, P=0.58; VE/Vco₂ slope >34, r=0.47, P=0.17). As demonstrated in Figure 1, FNDC5 expression correlated with PGC-1α expression in relation to the entire cohort (r=0.61 P=0.001). Correlations between PGC-1α and FNDC5 and peak Vo₂ and the VE/Vco₂ slope are illustrated in Figure 2. Although all correlations trended in the expected direction, only the relationship between PGC-1α and peak Vo₂ reached statistical significance.

BMI indicates body mass index; NT-proBNP, N-terminal prohormone of brain natriuretic peptide; ACE, angiotensin converting enzyme; ARB, angiotensin (II) receptor blocker; Vo₂, oxygen consumption; VE/Vco₂, minute ventilation/carbon dioxide production; NS, not significant; and NYHA, New York Heart Association.

LVEF indicates left ventricular ejection fraction; LVIdd, left ventricular internal dimension at diastole; FS, fractional shortening; PASP, pulmonary artery systolic pressure; MV, mitral valve; Vo₂, oxygen consumption; VE/Vco₂, minute ventilation/carbon dioxide production; RER, respiratory exchange ratio; and NS, not significant.
This is the first study to show that expression of FNDC5, the genetic forerunner to irisin, may impact aerobic performance in HF patients and help to broaden our understanding of the biological determinants of exercise capacity and related cardiovascular health. Traditionally, the symptoms and progression of HF have been attributed solely to cardiac dysfunction and remodeling. Our findings regarding FNDC5 are part of an expanding literature demonstrating that skeletal muscle and other peripheral tissues mediate HF symptoms and pathophysiology.

Although most HF patients lose muscle mass, leading to frailty and functional decline, the underlying mechanisms remain unclear. We demonstrate that expression in skeletal muscle of PGC-1α and FNDC5 are higher and correlate to one another in a more functional HF cohort, in contrast to the lack of correlation in those who are functionally limited. The recent identification of irisin, the product of the FNDC5 gene, suggests the presence of a hormonal pathway between muscle and adipose tissue, which may mediate some of the beneficial effects of exercise. Recent groundbreaking experiments in mice demonstrate that the transcriptional coactivator, PGC-1α, which promotes biogenesis of mitochondria, drives the expression of FNDC5, which in turn leads to increased brown adipose tissue, increased Vo2, insulin sensitivity, and glucose tolerance. Despite the known presence of irisin in human plasma, FNDC5 expression in the skeletal muscle of human systolic HF patients has not been previously reported.

Furthermore, our data are the first to correlate increased PGC-1α and FNDC5 expression in subjects with higher aerobic performance characteristics, as defined by key CPX indices. These findings in human muscle are consistent with the data of Boström et al in mice linking muscle expression of PGC-1α with FNDC5 expression. It seems likely that higher contractile activity in more aerobically fit HF patients promotes PGC-1α and FNDC5 expression. A sensitive assay for identification of the irisin protein in human plasma is not yet available so we were unable to directly measure irisin levels in this patient cohort.

Peak Vo2 is considered the cornerstone of functional evaluation in HF patients and is assessed for quantification of disease severity and prognosis. However, ventilatory inefficiency (ie, the VE/VCO2 slope) is also associated with increased risk for hospitalization and mortality through somewhat overlapping but different physiologic mechanisms. Although there are conflicting data regarding the optimal parameter for assessing functional capacity in HF patients, we show that regardless of the CPX parameter used for functional stratification, reduced performance is still associated with reduced PGC-1α and FNDC5 expression. Consistently, in the lower-functioning study population subgroups, PGC-1α expression did not correlate with FNDC5 expression, suggesting that the lack of signaling through these pathways may in part contribute to skeletal muscle

### Table 3. Gene Expression

<table>
<thead>
<tr>
<th></th>
<th>Entire Cohort (n=24)</th>
<th>Peak Vo2 (mL O2·kg⁻¹·min⁻¹) (n=15)</th>
<th>Peak Vo2 (mL O2·kg⁻¹·min⁻¹) (n=9)</th>
<th>VE/VCO2 Slope (n=14)</th>
<th>VE/VCO2 Slope (n=10)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>PGC1-α</td>
<td>7.9±5.4</td>
<td>9.9±5.9</td>
<td>4.5±1.9</td>
<td>&lt;0.01</td>
<td>9.7±6</td>
<td>5.3±3.4</td>
</tr>
<tr>
<td>FNDC5</td>
<td>4.7±3.11</td>
<td>5.7±3.5</td>
<td>3.1±1.2</td>
<td>&lt;0.05</td>
<td>5.8±3.6</td>
<td>3.3±1.4</td>
</tr>
</tbody>
</table>

PGC1-α and FNDC5 expression levels are given as mean±SD relative to the expression of glyceraldehyde 3-phosphate dehydrogenase performed in the same sample of cDNA.

Vo2 indicates oxygen consumption; VE/VCO2 slope, minute ventilation/carbon dioxide production.

![Figure 1. Correlation between FNDC5 and PGC-1α in the total study population. PGC1-α indicates peroxisome proliferator-activated receptor-γ coactivator-1α; FNDC5, α; and FNDC5, fibronectin type III domain containing 5.](image-url)
abnormalities and functional decline among HF patients whose disease severity is at an advanced stage.

Notably, only PGC-1α linearly correlated significantly with peak $V_{\text{O}_2}$ (Figure 2), and the relationship was only modest. This finding indicates that although dichotomous classification of the cohort according to established peak $V_{\text{O}_2}$ and VE/V$V_{\text{CO}_2}$ slope thresholds (as stratifications of disease severity) seem to identify differences in FNDC5 and PGC-1α expression, these same CPX variables do not seem to explain differences in FNDC5 and PGC-1α in a linear, continuous fashion. If confirmed by future investigations in larger cohorts, these findings may lead to recommendations regarding CPX data application in relation to skeletal muscle physiologic assessments, ie, specific CPX thresholds of high versus low performance may be useful as key categories relative to which skeletal muscle physiology can be better understood and characterized.

Boström et al also suggest that irisin administration may have therapeutic benefits on glucose tolerance and obesity that mimic or amplify effects of exercise. Irisin may have particular benefits in a systolic HF population in which comorbidities such as obesity and insulin resistance are often present. It is not yet known whether expression of FNDC5 is particularly low in these populations or if augmenting irisin levels in these patients will lead to clinical improvements. Although it is known that exercise can improve functional status in HF patients, it is likewise not yet known how FNDC5 expression changes with exercise training in these patients. Studies are currently underway to address these issues.

Limitations
A clear limitation of this study is the small sample size, but because the data reinforce and extend seminal principles published in recent animal studies it still stands out as an important addition to our understanding of irisin biology. Among the implications of a small study population, the degree of correlation between FNDC5 and PGC-1α expression and each of the CPX measurements may have been affected. Thus, although we demonstrated differences of gene expression in relation to categorizations of high-aerobic performance based on peak $V_{\text{O}_2}$ and VE/V$V_{\text{CO}_2}$ slope, it is not clear whether this reflects a physiologic distinction (such as disproportionate elevation of VE/V$V_{\text{CO}_2}$ slope despite similar peak $V_{\text{O}_2}$ values in 2 HF patients based on differences in pulmonary pressures).
or variance that is merely attributable to inadequate sampling. Additional research in larger cohorts is needed to further clarify these associations.

A related limitation is that a healthy control population was not included. While we considered comparing our findings with FNDC5 and PGC-1α expression to a cohort without HF, those without HF achieved much more favorable performance measures than HF patients (based on peak VO₂ and the VE/VCO₂ slope). Therefore in contrast to validated prognostic stratifications of peak VO₂ and the VE/VCO₂ slope for HF, it was unclear how to stratify the much higher performance results of the non-HF cohort and to make meaningful comparisons to gene expression in HF patients.

More fundamentally, we were limited to assessments of FNDC5 expression as assays for irisin are not yet available, but conclusions regarding irisin based on FNDC5 are still physiologically robust. An additional limiting factor is that we have only assessed correlations between PGC-1α and FNDC5 gene expression in functionally stratified HF patients and therefore cannot establish causality.

Influence of sex-specific variables, including hormonal influences, on gene expression and muscle metabolism may also be relevant. While we limited enrollment to male subjects to minimize confounding sex effects, evaluation of FNDC5 expression in females is needed in future investigations. Also, although there were no significant differences among patient demographic characteristics in the high-versus low-performance groups, this may be attributed to the small sample size than to true biological equivalence. N-terminal-pro B-type natriuretic peptide levels, for example, were substantially higher in the lower functioning groups (assessed by either peak VO₂ or the VE/VCO₂ slope). It is uncertain if this peptide is associated with or influences relative differences in PGC-1α and FNDC5 gene expression.

Conclusions

The present investigation demonstrates that PGC-1α and FNDC5 gene expression are increased in HF patients who have better aerobic performance, as indicated by the 2 primary CPX variables assessed in the HF population. Furthermore, PGC-1α correlates with FNDC5 only in the higher-functioning groups.

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Disclosures

None.

References


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**CLINICAL PERSPECTIVE**

Exercise intolerance is a common symptom among heart failure (HF) patients with profound clinical ramifications. Both central (cardiac output, alveolar perfusion) and peripheral (skeletal muscle, capillary perfusion) mechanisms have been implicated. Moderate-intensity aerobic exercise training typically improves functional capacity with little or no impact on central cardiac parameters, highlighting the importance of peripheral physiology on HF management. Nonetheless, pertinent HF effects on skeletal muscle remain poorly defined. Aerobic exercise training has been demonstrated to induce a more oxidative skeletal muscle phenotype, characterized by increased expression of the transcriptional coactivator peroxisome proliferator-activated receptor-γ coactivator-1α (PGC-1α) and mitochondrial growth. Recent animal studies have also demonstrated that irisin, a newly described myokine, is induced by exercise through the action of PGC-1α to stimulate changes in adipose tissue that mediate improvements in systemic metabolism. We hypothesized that expression of PGC-1α and FNDC5 (the genetic precursor of irisin) relates to aerobic performance in human HF patients. We measured aerobic capacity (cardiopulmonary exercise testing indices peak oxygen consumption and ventilatory efficiency slope) in 24 patients with systolic HF (left ventricular ejection fraction ≤40%). We assessed PGC-1α and FNDC5 from skeletal muscle biopsies. We demonstrate that PGC-1α and FNDC5 gene expressions are greater in the patients with the greater functional capacity (peak oxygen consumption >14 mL·kg⁻¹·min⁻¹, ventilatory efficiency slope <34). This is the first study to show that irisin expression correlates with functional capacity in HF patients. Lower FNDC5 expression may underlie reduced aerobic performance in HF.
Expression of the Irisin Precursor FNDC5 in Skeletal Muscle Correlates With Aerobic Exercise Performance in Patients With Heart Failure
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