Myocardial Expression Level of Neural Cell Adhesion Molecule Correlates With Reduced Left Ventricular Function in Human Cardiomyopathy

Kazuya Nagao, MD; Naoya Sowa, AD; Katsumi Inoue, MD; Motoko Tokunaga, MD; Kohei Fukuchi, MD; Koji Uchiyama, MD; Haruyasu Ito, MD; Fujio Hayashi, MD; Toshinori Makita, MD; Tsukasa Inada, MD; Masaru Tanaka, MD; Takeshi Kimura, MD; Koh Ono, MD

Background—Recently, we screened for cardiac genes induced by metabolic stress and identified neural cell adhesion molecule (NCAM) as a candidate. This study aimed to clarify the expression pattern of NCAM in human cardiomyopathy.

Methods and Results—A total of 64 cardiac tissue samples of patients with dilated cardiomyopathy were dichotomized according to the immunohistochemically determined signal intensity of NCAM staining (NCAM-high and NCAM-low groups). Clinical and hemodynamic data of the patients were compared between the 2 groups. Fibrosis area, left ventricular end-diastolic volume index, and left ventricular diastolic pressure were greater in the NCAM-high group (22.8% versus 11.6%, \( P<0.05 \); 130.3±57.6 versus 104.8±31.7 mL/m², \( P<0.05 \); 14.3±8.0 versus 8.8±4.7 mm Hg, \( P<0.005 \), respectively). Incidence of cardiac death and admission for worsening heart failure was higher in the NCAM-high group during a follow-up of 6.3 years (log-rank \( P<0.05 \)). Another 18 tissue samples were analyzed to determine the relationships between expression level of NCAM and major metabolic genes as well as hemodynamic parameters. The mRNA level of NCAM correlated with the serum \( (r=0.58; P=0.01) \) and mRNA levels \( (r=0.61; P=0.008) \) of brain-derived natriuretic peptides. It was also correlated with the mRNA levels of proliferator-activated receptor-\( \gamma \) coactivator-1 \( \alpha \) \( (r=0.69; P=0.002) \) and the nuclear respiratory factor 1 \( (r=0.74; P<0.001) \).

Conclusions—Expression of NCAM was associated with worsening hemodynamic parameters and major metabolic genes. Together with our previous findings, these data support the involvement of NCAM in left ventricular remodeling, revealing new insights into the pathophysiology of heart failure. (Circ Heart Fail. 2014;7:351-358.)

Key Words: cardiomyopathies ● heart failure ● neural cell adhesion molecule
area where expression of NCAM was closely associated with fibrotic change and then expanded throughout the myocardium during the advance of LV remodeling.14 These results indicate that NCAM may be involved in LV remodeling and have potential as a new therapeutic target or a biomarker in the clinical setting. Therefore, confirming the expression of NCAM in failing human hearts is of great significance.

Reports on the expression pattern of NCAM in failing human hearts are scarce, and it is currently unknown whether NCAM is also upregulated in the failing human heart as we observed in the animal model. Accordingly, we sought to investigate the expression pattern of NCAM in human LV tissue. We analyzed the relationships between NCAM expression levels and hemodynamic parameters, as well as the expression levels of major genes involved in metabolic regulation.

Methods

Research Design and Patient Population

We analyzed the expression of NCAM in the tissues of LV endomyocardial biopsies (EMBs) and examined the relationships between its level and hemodynamic parameters as well as other gene expression levels in patients with dilated cardiomyopathy (DCM). EMB was performed at Osaka Red Cross Hospital to determine the pathogenesis of HF, especially to rule out myocarditis and other specific myocardial diseases, according to the following institutional criteria: (1) new onset HF with clinical symptoms of dyspnea, chest pain, or palpitation, (2) unexplained impairment of LV function and LV dilation, (3) no evidence of coronary artery disease or primary valvular disease. These criteria were in agreement with a later published guideline on EMBs.29,30 Final diagnoses were obtained on the basis of clinical history, laboratory examinations, ECG, echocardiography, disease-specific tests such as computed tomographic scan and nuclear imaging, biopsies from extracardiac tissues, as well as histopathologic analysis of EMB tissue.29,30 This study is consisted of the following 2 groups of patients.

Immunohistochemical Analysis Group

Between March 2000 and July 2007, a total of 64 patients were diagnosed with DCM after undergoing EMB. We retrospectively examined EMB tissues obtained from these patients for immunohistochemical analysis. All patients underwent left heart catheterization, and 59 (92%) patients also underwent right heart catheterization. All patients provided written informed consent for the procedure and gene expression analyses. Because of the reduced renal function, all patients provided written informed consent for the procedure and gene expression analyses. The ethics committee of Osaka Red Cross Hospital approved the procedure and gene expression analyses. The ethics committee of Osaka Red Cross Hospital approved the procedure and gene expression analyses.

Gene Expression Analysis Group

We examined 18 EMB tissue samples obtained from patients with DCM who underwent EMB between February 2011 and October 2012 to analyze the relationships between gene expression level of NCAM and hemodynamic parameters. All patients underwent left heart catheterization, and 17 patients also underwent right heart catheterization. For 1 patient, we did not perform left ventriculography because of the reduced renal function. All patients provided written informed consent for the procedure and gene expression analyses. The ethics committee of Osaka Red Cross Hospital approved the study protocol.

Heart Catheterization and EMB

In right heart catheterization, pulmonary artery pressure, pulmonary artery wedge pressure, right ventricular pressure, right atrial pressure, and cardiac index were obtained using a triple-lumen Swan–Ganz catheter. In left heart catheterization, selective coronary angiography was performed to rule out coronary artery disease after aortic pressure recording. Thereafter, LV pressure recordings and left ventriculography were performed using a pigtail catheter to measure the LV end-diastolic pressure, LV systolic pressure, LV end-diastolic volume index, and LV end-systolic volume index. Finally, LV EMB was performed via the femoral artery using myocardial biopsy forceps (Technowood, Tokyo, Japan) after retrograde passage of the aortic valve. Tissues were obtained from the LV endomyocardium in each patient. No procedural complications occurred in the study population.

Immunohistochemistry

Immediately after being obtained, biopsy tissues were fixed in 10% buffered formalin and embedded in paraffin. Serial sections were obtained and stained with hematoxylin and eosin and Masson trichrome stain. When required, disease-specific analyses, such as Congo red or Gomori-trichrome staining, were also performed. The remaining paraffin blocks were stored for further immunohistochemistry. For histologic examination of NCAM, we obtained additional sections that were sequential with sections stained with Masson trichrome to evaluate the relationship between the expression pattern of NCAM and fibrotic change. Immunohistochemical staining of NCAM was performed using a conventional avidin–biotin complex method as described previously.14 To eliminate interpreparative variability in the immunohistochemical data, 10 sections were immunostained in a single batch using identical aliquots of diluted antibodies and other reagents.

Histopathologic Analysis

For each specimen, we selected the largest or most well-preserved tissue for histopathologic analysis. Images of each section were acquired using a microscope at a magnification of ×40, which covered almost the whole sample area. The percentage area of fibrosis was calculated by dividing the sum of the fibrotic areas of the section by that of the total tissue area as described previously.15 For semiquantitative analysis of NCAM staining, 3 areas cut in a plane parallel to the long axis of the cardiac myocytes were selected at a magnification of ×200 in each section, where staining of NCAM in intercalated discs was most clearly observed. The signal intensity of NCAM staining was analyzed in 10 randomly selected intercalated discs in each area using ImageJ software, and the average signal intensity of NCAM was calculated as NCAM intensity value in each specimen. Accordingly, we analyzed NCAM expression at 30 points per specimen. The details of the method of quantification are described in the results. When the intensity value of NCAM staining exceeded the median value (68.4 arbitrary units), the specimen was categorized into the NCAM-high group, whereas those categorized into the NCAM-low group had values below the median value. The analysis was performed in a blinded manner.

Follow-Up

Clinical follow-up was performed retrospectively in the patients included in the immunohistochemical analysis until March 2012 for a median duration of 6.3 years (interquartile range, 4.2–9.3 years) after EMB was performed. Clinical information was obtained either from a review of the medical records or by telephone or by letter contact with the patients or family members. Clinical follow-up information for the NCAM-high and the NCAM-low groups was obtained from 30 patients (93.8%) and 29 patients (90.6%) at 1 year, 28 patients (87.5%) and 28 patients (87.5%) at 3 years, and 16 patients (50.0%) and 23 patients (71.9%) at 6 years.

RNA Extraction and Quantitative Real-Time Polymerase Chain Reaction

Total RNA was isolated and purified using TRIzol reagent (Invitrogen, CA), and cDNA was synthesized from 1 μg of total RNA using a Transcript First Strand cDNA Synthesis Kit (Roche, Basel, Switzerland) in accordance with the manufacturer instructions. Quantitative real-time polymerase chain reaction (qRT-PCR) was performed using an SYBR Green PCR master mix (Applied Biosystems, CA), and the products were analyzed using a thermal cycler (ABI Prism 7900HT sequence detection system). The levels of β-actin transcripts were used to normalize cDNA levels. Gene-specific primers were as the following:

NCAM sense, 5′ GTGTTGTTACAGGCGAGGAT-3′

antisense, 5′ GATGACACCTCGGCTTTTG-3′

PGC-1α sense, 5′ TGACACAACCGGGACAA-3′

Downloaded from http://circheartfailure.ahajournals.org/ on August 30, 2017
NGO et al  NCAM Is Upregulated in Failing Human Heart 353

antisense, 5′ GCATCACAGGTATAACGGTAGG-3′
NRFI sense, 5′ GGCACTGTCTCACTTATCCAGTT-3′
antisense, 5′ CAGCCACGGCAGATAATTCA-3′
Tfam sense, 5′ AATGGATAGGCCAGGAAC-3′
antisense, 5′ CAAGTATTGTCTGAGAAGTC-3′
β-actin sense, 5′ AGGCACCAGGGCAGAT-3′
antisense, 5′ TGTCCCAGTTGACGAT-3′.

PGC-1α stands for proliferator-activated receptor gamma coactivator-1 α; NRF1, nuclear respiratory factor 1; and Tfam stands for mitochondrial transcription factor A.

Statistics
Frequency analysis was performed by χ2 test. Continuous variables were expressed as the mean value±SD or median with interquartile range. The differences in continuous variables between the 2 groups were assessed by Student t test or by nonparametric Mann–Whitney U test as appropriate. Survival rates were calculated using the Kaplan–Meier method and were compared using the log-rank test. Correlations were tested with the use of the Spearman correlation coefficient or Pearson correlation coefficient as appropriate. Probability values <0.05 were considered as statistically significant.

Results
Histopathologic Findings of Biopsy Specimens
To analyze the expression pattern of NCAM in human remodeling hearts, and especially its relationship with fibrosis, we performed immunostaining of NCAM and Masson trichrome staining in sequential sections of specimen from patients with DCM. In tissues where fibrotic change was scarce and normal-appearing cardiac myocytes remained intact, staining of NCAM was weak and restricted to the intercalated discs (Figure 1A). However, in sections where massive fibrotic change occurred, strong staining of NCAM was observed in intercalated discs and sarcolemma in cardiac myocytes adjacent to the fibrotic area (Figure 1B).

Semiquantitative Analysis for Expression Level of NCAM
A total of 64 patients with DCM were included in the immunohistochemical analysis. The mean ejection fraction was 40±13%.

Figure 1. A and B, Images of tissues with mild (A) and severe (B) fibrotic change obtained from dilated cardiomyopathy (DCM) patients. In each group, 2 representative images are shown. Left, Masson trichrome staining; middle, NCAM staining; right, high power field of the square area shown in the middle. Left and middle are sequential sections.
and the mean LV end-diastolic volume index (LVEDVI) was 118±48 mL/m² (Table 1). We performed semiquantitative analysis of the expression level of NCAM based on the digitally determined strength of NCAM immunoreactivity as described in the Methods and Figure 2. We categorized the 64 patients into 2 groups according to the median value of NCAM signal intensity; an NCAM-low group (median value = 68.4 arbitrary units [a.u.]) and an NCAM-high group (median value = 68.4 a.u.). There was no difference in clinical background or medications between the NCAM-low and NCAM-high groups (Table 2).

**Comparison of Fibrosis and Hemodynamic Parameters Between the 2 Groups**

The extent of the histologically determined fibrotic area was significantly larger in the NCAM-high group compared with the NCAM-low group (22.8% [interquartile range, 10.9–38.7%] versus 11.6% [interquartile range, 6.3–17.1%], P<0.05; Table 3). LVEDVI and LV diastolic pressure (LVDP) of patients categorized into the NCAM-high group were also significantly higher than those in the NCAM-low group (130.3±57.6 versus 104.8±31.7 mL/m², P<0.05; 14.3±8.0 versus 8.8±4.7 mm Hg, P<0.005, respectively). No difference was observed in ejection fraction, mean aortic pressure, mean pulmonary artery pressure, mean pulmonary capillary wedge pressure, or cardiac index between the 2 groups (Table 3).

**Table 1. Baseline Clinical Characteristics**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Patients, n 64</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>59±13</td>
</tr>
<tr>
<td>Men, n (%)</td>
<td>42 (66)</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>23±4</td>
</tr>
<tr>
<td>Diabetes mellitus, n (%)</td>
<td>8 (13)</td>
</tr>
<tr>
<td>Hypertension, n (%)</td>
<td>15 (23)</td>
</tr>
<tr>
<td>Dyslipidemia, n (%)</td>
<td>8 (13)</td>
</tr>
<tr>
<td>Left ventricular ejection fraction, %</td>
<td>39.8±12.9</td>
</tr>
<tr>
<td>Left ventricular end-diastolic volume index, mL/m²</td>
<td>117.6±47.9</td>
</tr>
<tr>
<td>Pulmonary capillary wedge pressure, mm Hg</td>
<td>10.9±5.7</td>
</tr>
<tr>
<td>Cardiac index, L/min per m²</td>
<td>2.9±0.9</td>
</tr>
<tr>
<td>Medication, n (%)</td>
<td>26 (41)</td>
</tr>
<tr>
<td>ACE inhibitor</td>
<td>25 (39)</td>
</tr>
<tr>
<td>ARB</td>
<td>12 (19)</td>
</tr>
<tr>
<td>β-Blocker</td>
<td>33 (52)</td>
</tr>
<tr>
<td>Furosemide</td>
<td>15 (23)</td>
</tr>
<tr>
<td>Spironolactone</td>
<td>9 (14)</td>
</tr>
<tr>
<td>Warfarin</td>
<td>13 (20)</td>
</tr>
<tr>
<td>Digoxin</td>
<td>6 (9)</td>
</tr>
<tr>
<td>Calcium channel blocker</td>
<td>2 (3)</td>
</tr>
</tbody>
</table>

Continuous variables are shown as mean±SD, and other values are n (%). ACE indicates angiotensin-converting enzyme; and ARB, angiotensin II receptor blocker.

**Clinical Follow-Up Outcomes**

During the follow-up period, there were 3 cardiac deaths, 5 noncardiac deaths, and 9 rehospitalizations for worsening HF. The cumulative incidence of all-cause death was not different between NCAM-low and NCAM-high groups (29.8% versus 6.9%, log-rank P=0.98; Figure 3). The cumulative incidence of adverse outcome defined by the combined end point of cardiac death and admission for worsening HF was significantly higher in the NCAM-high group compared with NCAM-low group at 1 year (19.2% versus 0%, log-rank P=0.01), at 3 years (22.4% versus 3.7%, log-rank P=0.03), at 6 years, (30.7% versus 7.6%, log-rank P=0.02), and in the overall cohort (31.5% versus 19.1%, log-rank P=0.03).
Table 2. Baseline Clinical Characteristics of Patients With DCM

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Low (n=32)</th>
<th>High (n=32)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>59±13</td>
<td>59±13</td>
<td>0.91</td>
</tr>
<tr>
<td>Men, n (%)</td>
<td>24 (75)</td>
<td>18 (56)</td>
<td>0.11</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>23±5</td>
<td>23±4</td>
<td>0.99</td>
</tr>
<tr>
<td>Diabetes mellitus, n (%)</td>
<td>6 (19)</td>
<td>2 (6)</td>
<td>0.13</td>
</tr>
<tr>
<td>Hypertension, n (%)</td>
<td>10 (31)</td>
<td>5 (16)</td>
<td>0.45</td>
</tr>
<tr>
<td>Dislipidemia, n (%)</td>
<td>5 (16)</td>
<td>3 (9)</td>
<td>0.45</td>
</tr>
<tr>
<td>Medication, n (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ACE inhibitor</td>
<td>14 (44)</td>
<td>11 (34)</td>
<td>0.44</td>
</tr>
<tr>
<td>ARB</td>
<td>15 (47)</td>
<td>11 (34)</td>
<td>0.31</td>
</tr>
<tr>
<td>β-Blocker</td>
<td>16 (50)</td>
<td>15 (47)</td>
<td>0.80</td>
</tr>
<tr>
<td>Furosemide</td>
<td>16 (50)</td>
<td>15 (47)</td>
<td>0.80</td>
</tr>
<tr>
<td>Spironolactone</td>
<td>17 (53)</td>
<td>16 (50)</td>
<td>0.80</td>
</tr>
<tr>
<td>Warfarin</td>
<td>6 (19)</td>
<td>6 (19)</td>
<td>1.00</td>
</tr>
<tr>
<td>Digoxin</td>
<td>10 (31)</td>
<td>5 (16)</td>
<td>0.14</td>
</tr>
<tr>
<td>Calcium channel blocker</td>
<td>7 (22)</td>
<td>2 (6)</td>
<td>0.07</td>
</tr>
<tr>
<td>Antiplatelet</td>
<td>9 (28)</td>
<td>4 (13)</td>
<td>0.12</td>
</tr>
<tr>
<td>Statin</td>
<td>3 (9)</td>
<td>3 (9)</td>
<td>1.00</td>
</tr>
<tr>
<td>Amiodarone</td>
<td>2 (6)</td>
<td>0 (0)</td>
<td>0.15</td>
</tr>
</tbody>
</table>

Continuous variables are shown as mean±SD, and other values are n (%). ACE indicates angiotensin-converting enzyme; ARB, angiotensin II receptor blocker; and DCM, dilated cardiomyopathy.

Relationship Between Expression of NCAM and Metabolic Genes

To elucidate whether upregulation of NCAM in human LV tissues was associated with response to metabolic stress, we analyzed the relationship between the expression of NCAM and genes involved in cardiac energy metabolisms. As shown in Figure 5A and 5B, the expression of NCAM was significantly correlated with those of PGC-1α, a major regulator of mitochondrial function and respiration in the heart (r=0.69, P<0.005), and its downstream genes, NRF1 (r=0.74, P<0.0005). No significant correlation was observed between the expression level of NCAM and transcription and mitochondrial maintenance factor (Figure 5C).

Discussion

In our previous report, we performed functional screening in combination with a signal sequence trap method and identified that NCAM was upregulated in cardiac myocytes under conditions of inhibition of mitochondrial ATP production. The mRNA levels of NCAM were also found to be remarkably upregulated by 24-fold during the remodeling period of hypertension-induced HF14; (2) more advanced LV remodeling was observed in the NCAM-high group as indicated by more severe fibrosis, larger LVEDVI, and higher LVDP compared with the NCAM-low group; (3) in accordance with these pathological and hemodynamic results, higher incidences of adverse cardiac events were observed in the NCAM-high group compared with the NCAM-low group; (4) upregulation of NCAM expression in cardiac tissue was associated with higher hemodynamic overload as indicated by increased level of circulating or expressing brain natriuretic peptides; (5) NCAM mRNA levels were correlated with the expression levels of metabolic genes such as PGC-1α and NRF1. These results support our previous work, suggesting the involvement of NCAM-mediated signaling in the pathophysiology of LV remodeling.

Table 3. Comparison of Fibrosis Area and Hemodynamic Parameters Between the NCAM-Low and NCAM-High Groups

<table>
<thead>
<tr>
<th>Histopathologic data</th>
<th>Low</th>
<th>High</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>% fibrosis</td>
<td>11.6 (6.3–17.1)</td>
<td>22.8 (10.9–38.7)</td>
<td>0.01</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Hemodynamic data</th>
<th>Low</th>
<th>High</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>LVEDVI, mL/m²</td>
<td>97.8 (82.4–123.8)</td>
<td>112.7 (94.7–148.7)</td>
<td>0.045</td>
</tr>
<tr>
<td>EF, %</td>
<td>40.0±11.2</td>
<td>39.7±14.6</td>
<td>0.95</td>
</tr>
<tr>
<td>LVEDP, mm Hg</td>
<td>8.5 (6.0–12.0)</td>
<td>13.0 (7.3–19.0)</td>
<td>0.001</td>
</tr>
<tr>
<td>PCWP, mm Hg</td>
<td>8.0 (5.0–11.0)</td>
<td>9.0 (6.0–17.0)</td>
<td>0.17</td>
</tr>
<tr>
<td>CI, L/min per m²</td>
<td>2.6 (2.2–3.4)</td>
<td>2.8 (2.2–3.3)</td>
<td>0.98</td>
</tr>
</tbody>
</table>

Continuous variables are shown as mean±SD or median (interquartile). CI indicates cardiac index; EF, ejection fraction; LVEDP, left ventricular end-diastolic pressure; LVEDVI, left ventricular end-diastolic volume index; NCAM, neural cell adhesion molecule; and PCWP, pulmonary capillary wedge pressure.
Reports concerning the expression pattern of NCAM in the heart are scarce. Gattenlöhner et al.32 screened for genes differentially expressed in human ischemic cardiomyopathy and identified NCAM as a candidate molecule. In their report, they demonstrated that NCAM was strongly expressed in cardiomyocytes adjacent to fibrotic areas in rat and human cardiac tissues, which was similar to our observation in DCM tissues. Therefore, a common upstream pathway for inducing expression of NCAM may exist in the perifibrosis area in ischemic cardiomyopathy and DCM. Arnett et al.33 recently performed a genome-wide association study of echocardiographic phenotypes in individuals with hypertension and identified NCAM as a genetic predictor of LV wall thickness and LV mass. In their report, they demonstrated that NCAM may be directly connected with numerous genes that have been shown to be involved with cardiac phenotypes. Taken together, expression of NCAM may be closely related to LV function and remodeling in a variety of human cardiac diseases.

Whether upregulation of NCAM in human LV tissue was triggered by metabolic stress consistent with our initial screening concept is of great interest. Because the energy demands in cardiac myocytes are high and must be precisely regulated, mechanisms to maintain energy homeostasis are known to exist.
PGC-1α has been shown to be a major regulator of mitochondrial function, biogenesis, and respiration in many tissues including the heart. Rohas et al demonstrated that under conditions of metabolic stress leading to decreased cellular ATP level, PGC-1α levels were induced remarkably and enhanced mitochondrial gene expression. This system worked as a compensatory system for maintaining cellular ATP level and cell survival.

In this context, increased expression of PGC-1α may indicate the existence of metabolic remodeling. Thus, we tried to determine the relationship between the expression levels of PGC-1α and NCAM. The results demonstrated that the expression level of NCAM was closely related with that of PGC-1α and its downstream target NRF1, suggesting that induction of NCAM expression might be associated with a compensatory response of PGC-1α during metabolic stress. Unfortunately, directly evaluating metabolic status, such as myocardial ATP content or the activity of metabolic enzymes, was limited by their instability and the small amounts of tissue available from LVEMB.

Ditlevsen et al showed that stimulation of NCAM-mediated signaling with a synthetic NCAM-ligand peptide protected neuronal cells against apoptosis, thereby showing the protective function of NCAM. Likewise, results in our previous experiments using NCAM-siRNA or pharmacological tools for NCAM stimulation demonstrated that NCAM-mediated signaling played a protective role under conditions of inhibition of mitochondrial ATP production in cardiac myocytes. Therefore, it would be reasonable to postulate that upregulation of NCAM expression in the human remodeling heart could be a compensatory response against cell death. In this regard, augmentation of NCAM-mediated signaling by recently developing NCAM-mimetic peptides may have therapeutic potential for cardiac diseases.

Future analysis using genetically manipulated animal models may clarify the function and mechanism of expression of NCAM in vivo in the failing heart.

Limitations
There are several limitations to the present study. First, the present study on the expression of NCAM in human LV tissues was based on the analysis of biopsy specimens, which represent regional molecular changes in the failing heart. Thus, the results of relationships between the molecular changes in sections and whole-heart hemodynamics may need careful interpretation. Second, we did not analyze biopsy specimens of normal human control hearts. However, variable degrees of LV dysfunction in the patients included in this study provided the opportunity to demonstrate the relationship between fibrosis, hemodynamic parameters, and NCAM expression. Third, concerning the relationships between the expression of NCAM and metabolic genes, we only analyzed the correlation of mRNA levels by qRT-PCR; therefore, we cannot establish the causality. Finally, our evaluation was limited by relatively small number of patients. In the cohort analysis, the low event rate precluded multivariate analysis. However, findings in human tissue demonstrated here reinforce and expand our previous observations in cells and animal models.

In conclusion, we showed for the first time that expression of NCAM was associated with worsening hemodynamic parameters and the expression of major metabolic genes in failing human hearts, as observed in the animal model. NCAM may be involved in LV remodeling, which provides new insights into pathophysiology of HF.

Acknowledgments
We thank Masayuki Shintaku for technical help.

Sources of Funding
This work was supported in part by grants from the Japan Society for the Promotion of Science; by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (to T. Kimura and K. Ono); by Grant-in-Aid for Scientific Research on Innovative Areas ‘Crosstalk between transcriptional control and energy pathways, mediated by hub metabolites’ (3307); and from the Ministry of Education, Culture, Sports, Science and Technology of Japan (to K. Ono).

Disclosures
None.

References


Myocardial Expression Level of Neural Cell Adhesion Molecule Correlates With Reduced Left Ventricular Function in Human Cardiomyopathy

Kazuya Nagao, Naoya Sowa, Katsumi Inoue, Motoko Tokunaga, Kohei Fukuchi, Koji Uchiyama, Haruyasu Ito, Fujio Hayashi, Toshinori Makita, Tsukasa Inada, Masaru Tanaka, Takeshi Kimura and Koh Ono

Circ Heart Fail. 2014;7:351-358; originally published online December 23, 2013; doi: 10.1161/CIRCHEARTFAILURE.113.000939

Circulation: Heart Failure is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX

Copyright © 2013 American Heart Association, Inc. All rights reserved.
Print ISSN: 1941-3289. Online ISSN: 1941-3297

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circheartfailure.ahajournals.org/content/7/2/351

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation: Heart Failure can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation: Heart Failure is online at:
http://circheartfailure.ahajournals.org//subscriptions/