Cardiac disease is a major national health concern with ≈15.4 million Americans having coronary heart disease and 5.1 million having heart failure, while incurring an annual cost of $300 billion.1 As interventions for acute cardiac events have evolved and become more effective, therapies aimed at mitigating the progression to heart failure have failed to keep pace. This void has led to the exploration of a variety of cell- and cytokine-driven vasculogenic and regenerative strategies.2–8 Preclinical studies of these approaches have yielded promising results, whereas early clinical trials have been encouraging but with more modest outcomes. As a result, it is necessary to build on these initial studies to enhance the translation to clinical therapeutics.

Background—Neuregulin-1β (NRG) is a member of the epidermal growth factor family possessing a critical role in cardiomyocyte development and proliferation. Systemic administration of NRG demonstrated efficacy in cardiomyopathy animal models, leading to clinical trials using daily NRG infusions. This approach is hindered by requiring daily infusions and off-target exposure. Therefore, this study aimed to encapsulate NRG in a hydrogel to be directly delivered to the myocardium, accomplishing sustained localized NRG delivery.

Methods and Results—NRG was encapsulated in hydrogel, and release over 14 days was confirmed by ELISA in vitro. Sprague-Dawley rats were used for cardiomyocyte isolation. Cells were stimulated by PBS, NRG, hydrogel, or NRG-hydrogel (NRG-HG) and evaluated for proliferation. Cardiomyocytes demonstrated EdU (5-ethyl-2'-deoxyuridine) and phosphorylated histone H3 positivity in the NRG-HG group only. For in vivo studies, 2-month-old mice (n=60) underwent left anterior descending coronary artery ligation and were randomized to the 4 treatment groups mentioned. Only NRG-HG–treated mice demonstrated phosphorylated histone H3 and Ki67 positivity along with decreased caspase-3 activity compared with all controls. NRG was detected in myocardium 6 days after injection without evidence of off-target exposure in NRG-HG animals. At 2 weeks, the NRG-HG group exhibited enhanced left ventricular ejection fraction, decreased left ventricular area, and augmented borderzone thickness.

Conclusions—Targeted and sustained delivery of NRG directly to the myocardial borderzone augments cardiomyocyte mitotic activity, decreases apoptosis, and greatly enhances left ventricular function in a model of ischemic cardiomyopathy. This novel approach to NRG administration avoids off-target exposure and represents a clinically translatable strategy in myocardial regenerative therapeutics. (Circ Heart Fail. 2014;7:619-626.)

Key Words: heart failure ■ ischemia ■ molecular biology ■ muscle cells ■ myocardial infarction

Cardiac disease is a major national health concern with ≈15.4 million Americans having coronary heart disease and 5.1 million having heart failure, while incurring an annual cost of $300 billion.1 As interventions for acute cardiac events have evolved and become more effective, therapies aimed at mitigating the progression to heart failure have failed to keep pace. This void has led to the exploration of a variety of cell- and cytokine-driven vasculogenic and regenerative strategies.2–8 Preclinical studies of these approaches have yielded promising results, whereas early clinical trials have been encouraging but with more modest outcomes. As a result, it is necessary to build on these initial studies to enhance the translation to clinical therapeutics.

Clinical Perspective on p 626

Neuregulin-1β (NRG) is a member of the epidermal growth factor family that plays a critical role in the development of neuronal, epithelial, and cardiac cells.9 It is a ligand to the ErbB4 tyrosine kinase receptor, activating an intracellular signaling cascade that leads to proliferation, differentiation, and migration.10 Importantly, the ErbB4 receptor is present in adult cardiomyocytes.11 The significance of NRG’s potential cardioprotective effects was highlighted in the clinical trials...
for trastuzumab, in which cardiac dysfunction was noted a
verse adverse effect.12 Trastuzumab is a monoclonal antibody that
inhibits the ErbB receptor and disrupts the NRG–ErbB sig-
naling pathway. More recently, it has been found that trastu-
zumab impairs human resident cardiac stem cells.13 These
findings support the notion that activation of the NRG–ErbB
pathway critically enhances cardioprotection.

From a therapeutic perspective, NRG has shown great prom-
ise in preserving cardiac function and inducing regeneration
after myocardial injury. This has been demonstrated in small
and large animal models14–18; however, multiple systemic intra-
venous infusions of NRG were required. This model of mul-
tiple parenteral infusions was translated to early clinical trial
in which a modest benefit in acute and chronic hemodynam-
ics was observed in patients with stable heart failure receiving
daily NRG infusions for 11 days.19 In addition, a phase II dou-
ble-blind study demonstrated improvements in left ventricular
(LV) ejection fraction and ventricular geometry using daily
infusions of NRG for 10 days.20 The clear drawback of sys-
temic NRG administration, however, is the significant potential
for adverse effects and the consequent dosing limitations.

To improve on this strategy, our group hypothesized that tar-
geted and sustained intramyocardial delivery of NRG would
yield NRG’s known cardioprotective and regenerative effects,
while avoiding systemic exposure and obviating the need for
multiple infusions. We sought to accomplish this by using an
established bioengineered hydrogel system21–24 to encapsulate
NRG for sustained and localized intramyocardial delivery in a
murine model of ischemic cardiomyopathy.

Methods

Macromer Synthesis
Sodium hyaluronate (74 kDa, Lifecore) was modified with hydroxyethyl
methacrylate (HEMA) to incorporate a terminal methacrylate group for
free-radical–initiated cross-linking and ester bonds to introduce hydro-
lytic degradation.25 Briefly, HEMA was reacted with succinic anhydride
via a ring opening polymerization in the presence of N-methylimidazole
to obtain HEMA-COOH, which was coupled to a tetrabutylammonium
salt of hyaluronic acid in the presence of 4-dimethylaminopyridine. The
resulting hyaluronic acid macromer with HEMA group modification
(HEMA–HA) was purified via dialysis, lyophilized, and the percentage of
HA disaccharides modified with an HEMA group was determined to be
≈15% using 1H nuclear magnetic resonance.

Hydrogel Gelation and NRG Encapsulation
With Release

To form hydrogels rapidly on injection into myocardium, a 2-com-
ponent redox initiator system consisting of ammonium persulfate
and N,N,N′-tetramethylethylenediamine was used. Ammonium
persulfate and N,N,N′-tetramethylethylenediamine were added to
a final concentration of 10 mMol/L in a 4% (wt/vol) HEMA-HA so-
lution and kept on ice. The kinetics of gel formation were character-
ized with rheometry at 37°C by monitoring the storage (G′) and loss
(G″) modulus with time, while applying oscillatory strain (20 mm
1° cone geometry, 1% strain, 1 Hz, Texas Instruments AR 2000ex).
Recombinant human NRG-1 (Thr176-Lys246) was acquired (R&D Systems,
Minneapolis) and used for the duration of the study in the NRG alone and NRG-HG
groups. For in vitro release kinetics, 2.5 μg NRG was added per 50
µL gel precursor solution, and 50 µL gels were formed in cylindrical
molds for 30 minutes at 37°C. Gels were incubated in 1 mL PBS sup-
plemented with 1% BSA at 37°C, and buffers were refreshed every 2
days. At t=0, 1, 2, 3, 5, 13, and 14 days, the supernatant was removed
and the released NRG was quantified by ELISA (Abcam, Cambridge,
MA) to determine the percentage of total NRG released. In parallel,
hydrogel degradation was quantified over 14 days with a uronic acid
assay as previously described.26

In Vitro Cardiomyocyte Isolation
Cardiomyocytes were isolated from neonatal Sprague-Dawley rat
pups using a trypsin-based dissociation method as previously de-
scribed,27 with minor modifications. The cell samples were centri-
fuged to wash away the trypsin solution and suspended in growth
media composed of: 4:1 ratio of DMEM and Medium-199, 10% horse
serum, 5% fetal bovine serum, 1% HEPES, and 1% solution of peni-
cillin, streptomycin, and glucose.

The cells were preplated onto multiple 10 mm2 petri dishes for 1
hour to allow fibroblasts to attach to the dishes. Cardiomyocytes still
suspended in the media were retained and counted. Cells were seeded
onto a glass bottom 24-well plate coated with vitronectin. The seed-
density for each well was 3.33×105 cells in 1 mL of growth media.
Growth media was changed 24 hours after seeding. Cells were kept in
culture for 5 days so that background proliferation would subside and
were randomized into 4 stimulation groups: PBS alone, NRG alone,
hydrogel (HG) alone, or NRG-HG. This time point is labeled as t=0
days. The media was changed daily. The hydrogels were kept suspen-
ded over the cells in a 3-μm pore size transwell cell culture insert. By
doing so, the daily medium changes did not wash away the hydrogel, which
could continue to release NRG into the fresh medium. In the NRG alone
group, the NRG was not replaced after the first medium change.

Cardiomyocyte Immunohistochemistry and
Confocal Microscopy
Cell proliferation was tested using immunohistochemistry at t=6 days.
Wells tested using the Click-iT EdU Imaging Kit (Life Technologies,
Carlsbad, CA) required incubation of the EdU reagent at t=4 days at a 1
µmol/L concentration. On t=6 days, all wells of the plate were washed
with PBS and fixed with 4% paraformaldehyde. Next, the cells were per-
meabilized using 0.5% Tween-20. Wells tested for EdU were incubated
with the detection solution and washed with rinse buffer. Cells were
also assayed for phosphorylated histone H3 (PH3). Cells were blocked
using 10% fetal bovine serum. Primary antibody for PH3 (Abcam,
Cambridge, MA) was added at 1:200. In addition, cells were stained
for cardiac troponin (Abcam) at 1:200. All antibodies were suspended
in PBS, and the cells were incubated in solution for 2 hours at 37°C.
After primary antibody incubation, the wells were washed with PBS
and the secondary antibodies (ab150065 and ab150112, Abcam) were
applied at 1:200 and incubated for 1 hour at 37°C. Lastly, the cells were
countereanstained for nuclei with DAPI (Life Technologies) and imaged
using a Zeiss LSM 710 confocal microscope (Oberkochen, Germany).

Animal Care and Biosafety
Male C57BL/6 mice weighing 25 to 30 g at 10 weeks of age were
obtained from Charles River Labs (Wilmington, MA). Food and wa-
ter were provided ad lib. All experiments conformed to the Guide
for the Care and Use of Laboratory Animals, published by the US
National Institutes of Health (eighth edition, 2011). The protocol was
approved by the Institutional Animal Use and Care Committee of the
University of Pennsylvania (protocol No. 803190).

Animal Model
Myocardial infarction was induced in 60 mice using an established
and highly reproducible model. Briefly, the mice were anesthetized
in a 2-L induction chamber (VetEquip, CA) and 3% isoflurane was
continuously delivered. A 20-gauge angiocatheter was used for
endotracheal intubation and connected to mechanical ventilation
(Hallowell EMC, MA) where 2% isoflurane was maintained through-
out. A thoracotomy was performed through the left fourth intercostal
space, the heart was exposed, and an 8-0 polypropylene suture was
placed around the left anterior descending artery 2 mm below the left
atrium producing an anterolateral myocardial infarction. The animals
were randomized into 4 groups (n=13/group) and received 3 peri-infarct intramyocardial injections of either PBS (40 μL), NRG alone (2.5 μg in 40 μL PBS), HG alone (40 μL), or NRG-HG (2.5 μg in 40 μL hydrogel). These borderzone injections were performed with the needle at a 20° angle to the myocardium to achieve an injection depth of 1 mm avoiding intraventricular delivery. The thoracotomy was closed, and tissue adhesive (VetBond, MN) was applied over the incision. All mice were implanted with subcutaneous microchips (BioMedic Data Systems, ID) for tracking and recovered from anesthesia. Buprenorphine (0.5 mg/kg) was administered for pain control.

**Echocardiographic Assessment**

LV geometry and function (n=32) were evaluated by echocardiography (Phillips, Amsterdam) at 2 weeks after myocardial infarction and therapy. LV parasternal short-axis 2-dimensional images were obtained at the papillary muscles. All analyses were performed by a single investigator who was blinded to the treatment groups.

**Immunohistochemistry and Histological Analysis**

At 6 days postoperatively, a subset (n=5/group) was anesthetized as previously described, and hearts were explanted. They were flushed with PBS, injected retrograde with Tissue Tek optimal cutting temperature compound (Netherlands), frozen at −80°C, and sectioned onto slides using a Leica CM3050S cryostat (Leica, Germany) at 10-μm thickness. Next, the samples were fixed with 4% paraformaldehyde and blocked with 10% fetal bovine serum. Sections were stained for PH3, Ki67, activated caspase-3, and NRG separately. All sections were stained for cardiac troponin. All primary antibodies (Abcam) were diluted at 1:200 in PBS and incubated for 2 hours at 37°C. Importantly, the NRG primary antibody (ab80237) was specific to the EGF-binding domain of human NRG to minimize cross-reactivity with mouse neuregulin. The sections were washed in PBS, and the appropriate secondary antibody was applied. The slides were washed in PBS and counterstained for nuclei with DAPI (4',6-diamidino-2-phenylindole; Vector Labs). The sections were imaged with a Leica DM5000B fluorescence microscope.

For histological analysis at 2 weeks, hearts were explanted, flushed with PBS, and injected retrograde with optimal cutting temperature compound through the aorta and pulmonary artery. Hearts were submerged in optimal cutting temperature compound, frozen, and stored in a −80°C freezer. Eight 10-μm-thick sections were prepared from each heart at the papillary level and stained with Masson trichrome. Standardized digital photographs were taken with a Nikon D5100 SLR camera (Nikon, Japan). Photographs were uploaded to ImageJ (version 1.46b), and the borderzone was assessed with digital planimetry.

**Peripheral Blood NRG Quantification**

At 6 days after myocardial infarction and therapy, a subset (n=5/group) was anesthetized, and 500 μL of peripheral blood was acquired. Blood was immediately placed in a biopsy tube with EDTA to prevent clotting and flash frozen. ELISA (ab100614, Abcam) was performed on all samples to quantify levels of human NRG. The samples were read on a microplate reader (TECAN, Austria) at 450 nm.

**Statistical Analysis**

All analyzed variables approximated a normal distribution, and values for continuous variables were reported as means±SD. For all experiments, a Levene test was first performed to establish homogenous variance across all groups. One-way ANOVA was then performed followed by a Tukey honest significant difference test to compare 2 groups at a time (α=0.05).

**Results**

**NRG Encapsulation and Sustained Release Kinetics**

NRG release was sustained for 14 days in vitro. An initial 2-day burst phase was observed where 60% of the NRG was released from the hydrogel. This was followed by a steady and sustained distribution of the remaining 40% of NRG for 12 days (Figure 1A). This release pattern was consistent with previous encapsulation of other similarly sized peptides. The uronic acid assay demonstrated 100% degradation of the hydrogel during a 14-day period as well, paralleling the release of NRG (Figure 1B).

**Sustained Release NRG Induces Cardiomyocyte Mitotic Activity in Vitro**

Isolated cardiomyocytes were stimulated with PBS, NRG alone, HG alone, or NRG-HG at t=0 days. Importantly, transwell plates that allowed passage of NRG released from hydrogel were used, and the media were changed daily. This allowed comparison of a 1-time administration of NRG versus sustained delivery. At t=6 days, the cardiomyocytes were visualized with confocal microscopy to evaluate for EdU incorporation, marking DNA synthesis. The PBS, NRG alone, and HG alone groups demonstrated no EdU incorporation compared with 10% of the cardiomyocytes in the NRG-HG group that did incorporate EdU (Figure 2A). Providing further evidence of cardiomyocyte cell cycle activation, immunohistochemistry was performed to determine the presence of PH3. There was no PH3 observed in the PBS, NRG alone, and HG alone groups, whereas the NRG-HG group exhibited PH3 positivity (Figure 2B) in 2% of the cardiomyocytes.

**Sustained Release NRG Induces Cardiomyocyte Mitotic Activity in Vivo**

At 6 days postoperatively, hearts were explanted for immunohistochemistry (n=5/group) and determination of the presence...
of PH3 and Ki67. Heart sections in the 3 control groups did not demonstrate evidence of PH3 or Ki67. In all of the NRG-HG–treated animals, heart sections were positive for PH3 and Ki67 (Figure 3A and 3B).

Sustained Release NRG Provides Cardioprotection In Vivo

In the same subset at 6 days postsurgery, heart sections (n=5/group) were evaluated by immunohistochemistry for the presence of activated caspase-3, a mediator of apoptosis and parameter of cardioprotection.28 This was quantified by number of positive cells per 10× high-powered field. The PBS (5.6±0.8 cells; P=0.005) and HG (6.2±0.97 cells; P=0.006) groups demonstrated significantly higher levels of activated caspase-3 compared with the NRG-HG group (1.5±0.29 cells; Figure 4). The NRG group demonstrated a greater number of positive cells (3.5±0.29 cells) compared with NRG-HG, but was not statistically significant.

Encapsulated NRG in Hydrogel Provides Sustained Local Release in Vivo Without Peripheral Exposure

In the same subset at 6 days postsurgery, peripheral blood (n=5/group) was acquired along with explanted hearts. Immunohistochemistry of explanted hearts demonstrated the

Figure 3. Mitotic activity in peri-infarct region. Heart sections at 6 days after treatment visualized under fluorescent microscopy demonstrating phosphorylated histone H3 (PH3) and Ki67 positivity with neuregulin (NRG)-hydrogel (HG) treatment. Sections are stained with DAPI (blue), troponin (red), PH3 (A) and Ki67 (B; green). All animals treated with NRG-HG demonstrated presence of PH3 and Ki67 in the peri-infarct region.
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presence of NRG in the myocardium in the NRG-HG group only (Figure 5A). The peripheral blood was assessed for the presence of NRG. There was no difference in peripheral blood NRG levels throughout all groups: native (3.6±0.4 ng/mL; \( P > 0.5 \)), NRG (3.4±0.3 ng/mL; \( P > 0.5 \)), HG (3.1±0.2 ng/mL; \( P > 0.5 \)), NRG-HG (3.5±0.2 ng/mL; Figure 5B).

Echocardiographic Assessment
At 14 days postoperatively, the animals were anesthetized and were evaluated by echocardiography (n=8/group). LV chamber area at the level of the papillary muscles was analyzed. The PBS (0.29±0.02 cm²; \( P = 0.05 \)), NRG alone (0.33±0.04 cm²; \( P = 0.02 \)), and HG alone (0.26±0.05 cm²; \( P = 0.03 \)) animals all demonstrated significantly greater LV chamber areas compared with NRG-HG–treated animals (0.18±0.02 cm²; Figure 6A). In addition, LV ejection fraction was determined at 14 days. The PBS (20.2±3.2%; \( P < 0.01 \)), NRG alone (20.1±4.7%; \( P < 0.01 \)), and HG alone (17.2±3.7%; \( P < 0.01 \)) groups all exhibited significantly reduced LV ejection fraction compared with NRG-HG–treated animals (37.5±3.6%; Figure 6B).

Histological Evaluation
Directly after echocardiography evaluation at 14 days, hearts were explanted and frozen sections were acquired for histological analysis (n=8/group). Masson trichrome staining was performed to evaluate borderzone thickness. The PBS (0.27±0.04 mm; \( P < 0.01 \)), NRG alone (0.30±0.05 mm; \( P < 0.01 \)), and HG alone (0.26±0.05 mm; \( P < 0.01 \)) groups revealed a significantly reduced borderzone thickness compared with animals receiving NRG-HG (0.65±0.07 mm; Figure 7).

Discussion
This study demonstrated that localized and sustained myocardial delivery of NRG stimulates cardiomyocyte mitotic activity, provides cardioprotection, reduces LV dilation, and enhances ventricular function in a murine model of ischemic cardiomyopathy. The novel and integral component of this study was the encapsulation of NRG in the hydrogel delivery system. Although previous preclinical and clinical studies have revealed a functional benefit of repeated systemic dosing of NRG, there remains a legitimate concern regarding NRG’s potential oncological consequences and its effect on the nervous system. The locally administered encapsulated NRG addresses this problem, allows for greater flexibility in dosing, and obviates the requirement for daily infusions. A crucial component of this study was confirming the presence of the administered NRG in the myocardium 6 days after injection.
only when delivered in a hydrogel. In addition, analysis of the peripheral blood at this time point exhibited no difference in NRG levels throughout all groups, including native mice. The low levels of NRG detected were probably attributable to minimal cross-reactivity between the detection antibody and native mouse neuregulin. These findings strongly support the strategy of sustained and local NRG delivery to the myocardium with minimal off-target exposure.

During the past decade, a multitude of bioengineered systems have been developed as adjuncts to cytokine, peptide, and cell therapies for heart failure. These include, but are not limited to, cell sheets, fibrin patches, alginate scaffolds, and hydrogels. The hyaluronic acid hydrogel was used in this study because of its previous effectiveness in encapsulating similarly sized peptides and its demonstrated safety in humans. There is strong evidence to suggest that administration of hydrogel alone contributes to cardiac repair and improvement in function; however, this was not observed in this study. The most logical explanation is that a very small volume of 40 μL was injected into the myocardium. This volume was optimal for encapsulating the determined dose of 2.5 μg of NRG and treating the borderzone.

After encapsulation of NRG, it was critical to determine that NRG could be released from the gel for an extended period of time without hindering its function. This was accomplished by incubating the NRG-containing hydrogel in PBS and evaluating the supernatant by ELISA for 14 days. The typical burst phase was observed followed by a steady release of NRG. After this analysis, in vitro experiments on isolated cardiomyocytes demonstrated that the released NRG retained its ability to activate the ErbB signaling axis. Importantly, these experiments were carefully performed such that the cells were cultured on a transwell plate where the hydrogel was suspended above the cells. This allowed for daily medium changes while keeping the hydrogel in place and continually releasing NRG into the new medium. In the NRG alone group, the NRG was not replaced after the first medium change. This allowed comparison between cardiomyocytes undergoing sustained NRG stimulation and a 1-time stimulation. The data demonstrated that activation of the cardiomyocyte cell cycle occurred only with sustained NRG administration. The NRG dosing was based on a previous study that characterized differentiated cardiomyocyte cell cycle re-entry after multiple days of NRG stimulation, which was consistent with the findings of this study. The visualization of EdU incorporation and PH3 positivity in NRG-HG–stimulated cells strongly supports that the integrity of the NRG–ErbB signaling axis was maintained.

After demonstration that NRG retains activity after hydrogel release, findings of reduced LV dilation and significantly improved LV ejection fraction coupled with increased borderzone thickness on histology 2 weeks after therapy illustrate enhancement of ventricular function and structure resulting from NRG-HG therapy. The in vitro and in vivo data exhibiting cardiomyocyte mitotic activity support the notion that NRG-HG–induced cardiomyocyte regeneration contributed to enhanced ventricular function. The decreased levels of activated caspase-3 after NRG-HG treatment also suggest an anti-apoptotic effect. It is logical that NRG administration would yield a protective effect in conjunction with a regenerative stimulus as the downstream pathway of the ErbB receptors includes PI3K (phosphoinositide 3-kinase) and Akt, which are well-established mediators of cellular proliferation and

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**Figure 6.** Echocardiography. Graphs demonstrating significantly decreased left ventricular (LV) chamber area at the papillary level (A) and significantly enhanced LV ejection fraction (EF; B) in the neuregulin (NRG)-hydrogel (HG) group compared with all controls. Measurements were determined by short-axis view on ECHO (*P<0.05 for each group vs NRG-HG).
In addition, the role of cardiac stem cells and NRG’s interaction occurred with already differentiated cardiomyocytes. Previous work has somewhat elucidated this mechanism in which fate mapping experiments showed that NRG-induced cardiomyocyte regeneration occurred with already differentiated cardiomyocytes. There were limitations to the study. Although the experiments confirmed the presence of NRG in the myocardium 6 days after injection, further information concerning the kinetics of encapsulated NRG administration would be helpful. This could be accomplished in future experiments by conjugating a fluorophore to NRG and using serial optical imaging to track its location over time. Regarding limitations involving delivery, intramyocardial injection poses a manageable hurdle when translating to clinical application. In addition to being an adjunct to cardiac surgery, hydrogels that are more suitable for catheter-based administration are being explored. As a result, the translational potential of this therapy is greatly broadened.

Overall, this study demonstrated that targeted and sustained delivery of NRG to borderzone myocardium stimulates cardiomyocyte mitotic activity and cardioprotection and significantly enhances ventricular function and structure. It is an extremely translatable therapy in that it addresses a major national health concern, NRG alone has reached clinical trial

Sustained and Targeted Neuregulin Delivery

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Disclosures

None.

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**CLINICAL PERSPECTIVE**

The central contribution of this study was the construction of a hydrogel delivery system to enable targeted and sustained intramyocardial administration of neuregulin-β1 (NRG) after myocardial infarction, leading to myocardial repair and enhanced ventricular function in a murine model of ischemic cardiomyopathy. The clinical impact may be significant in that systemic administration of NRG for patients with heart failure has reached clinical trial, which greatly augments the translatability of this targeted approach. A major advantage of localized and sustained delivery of NRG is that it avoids off-target exposure, thereby allowing more flexibility in future dosing. In addition, as demonstrated by this study, sustained NRG stimulation is necessary to induce myocardial regeneration and enhance function. As hydrogel-encapsulated NRG could be delivered via a catheter-based endovascular approach or an open surgical approach, the potential clinical applications are significant and broad. Preclinical large animal studies are currently underway to examine these strategies and evaluate targeted NRG delivery for potential clinical translation.
A Bioengineered Hydrogel System Enables Targeted and Sustained Intramyocardial Delivery of Neuregulin, Activating the Cardiomyocyte Cell Cycle and Enhancing Ventricular Function in a Murine Model of Ischemic Cardiomyopathy


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