Myocardial infarction (MI) leads to molecular, structural, geometric, and functional changes of heart in a process known as post-infarction cardiac remodeling. Despite modern reperfusion strategies, patients who survive an acute MI are at higher mortality rate because of tonic sympathetic activities that promote ventricular remodeling. It is well known that chronic vagus nerve stimulation (VNS) could suppress the cardiac sympathetic outflow and prevent and reverse cardiac remodeling. However, VNS treatment has to implant neurostimulator system with a bipolar multicontact cuff electrode around the cervical vagus nerve by surgery and may cause side effects, including neck pain, coughing, swallowing difficulty, and voice alteration along with nausea and indigestion. Transcutaneous electric stimulation of auricular branch of vagus nerve (ABVN) located at the tragus, the anterior protuberance of the outer ear, is a noninvasive approach to stimulate afferent vagus nerve fibers. Our previous research found low-level tragus stimulation (LL-TS) could substitute for VNS to reverse acute atrial remodeling. Nevertheless, there was no further research about whether LL-TS could affect ventricular remodeling after MI. The present study was designed to test the hypothesis that chronic intermittent LL-TS could attenuate ventricular remodeling in conscious dogs with healed MI.

Methods

Animal Preparation

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The protocol was approved by the Committee on the Ethics of Animal Experiments of the...
Wuhan University (Permit Number: 2014-0363). All surgery was performed under sodium pentobarbital anesthesia, and all efforts were made to minimize suffering. Thirty adult male beagle dogs (body weight, 10–15 kg) were supplied by the center of experimental animal in medical college of Wuhan University. All dogs were anesthetized with Na-pentobarbital, 40 mg/kg, and ventilated with room air by a positive pressure respirator (MAA01746, Harvard Apparatus Holliston, MA). Additional maintenance doses of 2 mg/kg Na-pentobarbital were administered at the end of each hour during the procedure. Normal saline at 50 to 100 mL/h was infused to replace spontaneous fluid losses. Body surface ECG was recorded by using subcutaneous needle electrodes during the whole procedure using a computer-based Laboratory System (Lead 2000B, Jinjiang Inc, China). A heating pad was used to maintain the core body temperature at 36.5±1.5°C. The thoracotomy was performed in the left fifth intercostal space. A Harris 2-stage occlusion was performed on the left anterior descending coronary artery with 3-0 silk above the first diagonal branch to produce MI and all the major diagonal branches were ligated to decrease collateral flow to the infarct area. The vessel was partially occluded for 20 minutes and then tied off completely. The left anterior descending artery occlusion was observed in ECG with elevation of ST segment after surgery. All parameters were measured 3× and evaluated blind to the intervention by the same experienced echocardiographer.

**Low-Level Tragus Stimulation**

Tragus stimulation (auricular VNS) as previously used (frequency 20 Hz, pulse width 1 ms) with duty cycle of 5 s on and 5 s off was delivered to the bilateral tragus in the external auditory canal with ear-clips connected to a custom-made stimulator (Figure 1). At first, incremental voltages were applied to the tragus until slowing of the sinus rate. The voltage necessary to slow the sinus rate was used as the threshold and LL-TS was delivered at 80% below that. The actual electric voltage was in the range of 16 to 24 V, which did not cause any heart rate changes and resulted in serious adverse reactions that could increase plasma epinephrine level.

**Experiment Protocol**

Thirty beagle dogs were randomly divided into 3 groups before surgery. MI group (n=10) was established by the ligation of left anterior descending artery and major diagonal branches to introduce MI. LL-TS group (n=10) underwent the same surgery with 4 hours of LL-TS at 7–9 AM and 4–6 PM that were highest sympathetic tone period proved by Irwin JM every day. Control group (n=10) underwent sham surgery without stimulation. All dogs were followed ≤90 days. None received other background therapy.

**Echocardiographic Evaluation of Left Ventricular Function**

Doppler echocardiography (Vivid E9, GE Healthcare) was performed under continuous ECG monitoring with a 3.5 MHz electronic probe on conscious dogs lying in lateral recumbency on the day before surgery, 30th day and 90th day after surgery, respectively. Images were obtained according to echocardiographic criteria and recorded in computer for subsequent analysis. The endocardial borders of left ventricle (LV) were measured at the bidimensional right long axis view. The LV end-systolic volume, LV end-diastolic volume, and LV ejection fraction were calculated using the Simpson’s biplane equation. The maximal left atrial volume (LA-Vma) at the standard left apical 4-chamber view was calculated using the Simpson’s monoplane method. The velocity waveforms of the peak mitral inflow velocity in early diastole (E) and during left atrial contraction (A) were measured to calculate the ratio of E to A. Systolic blood pressure, which was measured on the left forelimb by the Ultrasonic Doppler Flow Detector (Model 811-B, Parks Medical, OR) and heart rate was collected during the echocardiographic examination on the day before the surgery and 90th day after surgery. All parameters were measured ≥3× and evaluated blind to the intervention by the same experienced echocardiographer.

**Myocardial Infarct Size Determination**

The infarct size was assessed with 0.5% Evans Blue and 1.0% triphenyltetrazolium chloride staining in 5 dogs of LL-TS group and 5 dogs of MI group. In brief, Evans Blue was infused into the left atrial appendage to evaluate the area at risk (AAR) by left thoracotomy under general anesthesia. After giving dogs the euthanasia, the heart was quickly removed and frozen at −20°C for 3 hours and then was cut into 2 to 5 mm thick slices perpendicular to the left anterior descending artery occlusion site. Five continuous slices from the occlusion site were incubated in triphenyltetrazolium chloride at 37°C for 15 minutes to discriminate the infarct tissues from the viable myocardium. After overnight fixation with 4% paraformaldehyde, each slice was photographed with a digital camera. The area measurement was performed using Image Tool software version 3.0. The ratio of the AAR to the total ventricular mass and the infarct size normalized to the AAR was calculated.6

**Histomorphometric Measurements**

At the end of protocol, after the dogs being euthanized, the hearts (5 dogs in LL-TS group and 5 dogs in MI group) were removed quickly from the chest for sectioning from apex to base into 3 transverse rings of 5 mm in thickness. Transmural tissue blocks that were obtained from infarct border zones of the middle slice were mounted on cork using Tissue-Tek embedding medium, and rapidly frozen in isopentane, precooled in liquid nitrogen. Cryostat sections ≈8 μm thick were prepared and stained with fluorescein-labeled peanut agglutinin (Vector Laboratories Inc) and used to delineate the myocyte border and the interstitial space, as previously described.7 Sections were double stained with rhodamine-labeled Griffonia simplicifolia lectin I to identify capillaries. Microscopic fields (magnification×100) were selected at random from each section and used to measure myocyte cross-sectional area. The surface area occupied by interstitial space and capillaries were measured from each field by means of computer-based video densitometry (Jandel Scientific, Corte Madera, CA) as previously described.8 For each histomorphometric measure, the sections obtained from each dog were averaged and that single
average was used to represent each dog in the analysis. The volume fraction of replacement fibrosis, namely, the proportion of scar tissue to viable tissue, was calculated from trichrome-stained sections as the percent total surface area occupied by fibrosis. The volume fraction of interstitial fibrosis was calculated as the percent total surface area occupied by interstitial space minus the percent total area occupied by capillaries.15

### Western Blotting

The hearts of 10 dogs from control group and the hearts that were sectioned in histomorphometric measurements were used for Western blotting. Transmural myocardial tissue sample ≈1 cm² obtained from the LV free wall outside the infarction area was homogenized in radioimmunoprecipitation assay lysis buffer (Rockland, Gilbertsville, PA) containing proteinase inhibitor (Roche, Basel, Switzerland). The homogenate was centrifuged at 4°C at 2000g for 10 minutes, and the resultant supernatant was further subjected to centrifugation at 12000g for 20 minutes. Protein concentration of each supernatant sample was determined using a DC Protein assay kit (BioRad, Hercules, CA). Samples containing equal amounts of protein were separated on 15% sodium dodecyl sulfate polyacrylamide gel electrophoresis gel (Bio-Rad) and transferred onto Immobilon-P membrane (Millipore, Billerica, MA). After blocking the membranes with BlockAce (Dainippon Pharmaceutical, Japan), the primary antibodies used in the study included polyclonal antibody for matrix metalloproteinase 9 (MMP-9; TheBinding Site, Birmingham, UK) and transforming growth factor β1 (TGF-β1), anti-collagen I, anti-collagen III (Abcam, Cambridgeshire, England), and β-Actin (Santa Cruz Biotechnology) which served as an internal control. The concentration of primary antibody was obtained from the manufacturer’s instructions. Detection was done with enhanced chemiluminescence (Invitrogen). Bands were scanned and analyzed on the ECL detection system (GE Healthcare).12 Experiments were performed in triplicate and repeated ≥3x.

### Blood Sampling

Blood samples collection was performed in a dedicated conscious testing room, where dogs were lying on a couch in a quiet, dim-lit environment, without any sedation or anesthesia. After a stabilization period of 15 minutes, venous blood samples were collected in ice-chilled tubes coated with EDTA (BD Vacutainer K2E, BD Diagnostics, NJ) on the day before the surgery and the 1st, 30th, and 90th day after surgery. Plasma was separated by centrifugation at 3000 rpm for 15 minutes at 4°C and stored at −80°C until assayed. Plasma high-specific C-reactive protein (hs-CRP) level was measured using a canine-specific high sensitivity CRP ELISA (KT-093, Kamiya Biomedical Company, Seattle), Plasma norepinephrine (NE), N-terminal pro-B-type-natriuretic peptide (NT-proBNP) concentrations were determined by a validated radioimmunoassay method supplied by American Laboratory Products Company (ALPCO, Salem). Plasma was processed in radioimmunoassay method according to the manufacturer’s published procedure.

### Data Analysis

All data were reported using the median with 25th and 75th percentiles except for the data in the Table. If significance was attained by Kruskal–Wallis test between 3 groups, post hoc pairwise comparisons with Dunn’s multiple comparison test was performed. Mann–Whitney nonparametric tests were used for comparison of fibrosis measurements (volume fraction of replacement fibrosis and volume fraction of interstitial fibrosis). The data in Table were analyzed using Wilcoxon signed-rank test (stroke volume and cardiac output) and paired t test (Weight, heart rate, and systolic blood pressure). For box plots in figures, the lower and upper bounds of the boxes indicate the 25th and 75th percentile (Q.25/Q.75) values, and the horizontal lines indicate the median. GrapPad Prism 6 for Mac (GrapPad Software Inc, La Jolla, CA) was used for statistical analysis. A value of \( P<0.05 \) was required for statistical significance.

### Results

**Effects of LL-TS on LV Function and Myocardial Infarct Size**

All dogs completed the experimental protocol. As was shown in Figure 2, the baseline of echocardiographic measurement was comparable in 3 groups. The LV end-diastolic volume and LV end-systolic volume were significantly higher and LV ejection fraction and E/A were significantly lower in MI group than in control group on the 30th and 90th day, and the LAV\(_{\text{max}}\) was significantly higher in MI group than in the control group on the 90th day. But LL-TS group significantly reduced LV dilatation, improved LV contractile, and diastolic function compared with MI group on the 30th and 90th day of follow-up. Besides, the treatment of LL-TS reduced the LA dilatation on the 90th day (all \( P<0.01 \)). However, as was shown in the Table, there were no significant differences in stroke volume and cardiac output at the end of 90-day follow-up between the 3 groups. Compared with baseline, dogs in MI group had significantly lower systolic blood pressure and higher heart rate on the 90th day (both \( P<0.01 \)), but the differences among 3 groups did not reach significance. Interestingly, a significant increase in body weight of all dogs was observed at the end of the 90-day follow-up (all \( P<0.01 \)), but there was no significant difference among the 3 groups.

Myocardial infarct size expressed as the percentage of AAR was shown in Figure 3 upside. The AAR, expressed as a percentage of the total ventricular mass, was a little larger in MI group than LL-TS group (M: median=46.2% [Q.25/Q.75=42.3%/49.7%] versus S: median=43.8% [Q.25/Q.75=40.1%/44.4%]; \( P=0.16 \)). However, LL-TS treatment significantly reduced mean infarct area by ≥50% compared
with MI group (M: median=48.6% [Q.25/Q.75=42.3%/49.7%] versus S: median=18.9% [Q.25/Q.75=17.1%/19.7%]).

**Effects of LL-TS on LV Remodeling**

Histomorphometric findings were shown in Figure 4. Treatments with LL-TS significantly decreased volume fractions of replacement fibrosis and interstitial fibrosis in the tissue of infarct border zones compared with dogs in MI group (all P<0.01).

The results of the determination of protein levels in LV free wall tissue are shown in Figure 5; the protein level of TGF-β1, MMP-9, collagen I, and collagen III was remarkably increased in MI and LL-TS group than in control group (all P<0.01). However, LL-TS group significantly reduced...
TGF-β1, MMP-9, collagen I, and collagen III protein expression level compared with MI group at the end of follow-up (all \(P<0.05\)).

Effects of LL-TS on Plasma hs-CRP, NE, and NT-proBNP Levels

Plasma levels of hs-CRP increased significantly on the first day after MI; LL-TS therapy markedly attenuated the increase trend on the 30th and 90th day (Figure 6, upper panel; both \(P<0.05\)).

Plasma NE levels also increased from the next day after induction of MI to the 90th day. However, the plasma levels of NE were significantly lower in LL-TS group than in MI group (Figure 6, middle panel; all \(P<0.05\)).

Plasma NT-proBNP levels were obviously increased from the 1st day to the 90th day after MI. However, LL-TS significantly reduced plasma NT-proBNP levels compared with MI group (Figure 6, bottom panel; all \(P<0.05\)).

Discussion

Major Findings

Low-level transcutaneous electric stimulation of ABVN has important impaction on cardiac function and ventricular remodeling. We found that chronic intermittent LL-TS treatment significantly improved cardiac function, alleviated cardiac fibrosis, and attenuated LV remodeling in conscious dogs with healed MI. Moreover, LL-TS treatment could also reduce plasma level of hs-CRP, NE, and NT-proBNP in dogs after MI.

Improvement of LV Remodeling by LL-TS Treatment

The improvement in global LV remodeling after MI was achieved by LL-TS treatment evidenced by reduced LA and LV dilatation and increased LV systolic and diastolic function. It has been proved that increased vagus nerve activity could reduce the ratio of infarct size to AAR by regulating the nicotinic action.\(^\text{13}\) Recently, Shinlapawittayatorn K et al\(^\text{14}\) also found that increased vagus nerve activity could reduce mitochondrial ROS production and mitochondrial swelling, which were responsible for decreasing the infarct size after MI. Our study also indicated the reduction of MI size was achieved by chronic intermittent LL-TS treatment. It probably was by means of enhancing vagus nerve activity that LL-TS improved the global LV remodeling.

Reversal of LV cellular remodeling by LL-TS treatment was shown with the reduction of LV fibrosis evidenced by the
metalloproteinase that inhibits MMP-9 activity. MMP-9 degrades collagen and contributes to ventricular dilation and remodeling in remote infarction areas after MI. This study found that the protein content of MMP-9 was markedly low on the 90th day after MI and LL-TS could further lower the MMP-9 level in LV tissue.

However, the exact mechanism of regulation of collagen expression level by LL-TS is still not clear. Maybe the renin–angiotensin–aldosterone system plays an important role in that, more study should be done to find out about it.

**Possible Mechanisms of LL-TS on Vagus Nerves**

The ABVN, which is the only peripheral branch of the vagus nerve, mainly supplies the auricular concha and most of the area around the auditory meatus. The afferent vagal nerve fibers innervating the tragus area enter the main vagal trunk through the jugular ganglion at level of the skull base. Nomura and Mizuno found that the afferent fibers of ABVN terminated mainly in the nucleus tractus solitaries. It is well known that a large number of autonomic nerve fibers, including the fibers from the heart, project to the nucleus tractus solitaries. Gao et al found acupuncture-like stimulation of ABVN evoked cardiovascular inhibition via activating the cardiac-related neurons in the nucleus tractus solitaries. Our previous researches pointed out that tragus stimulation activated a series of neurotransmission between ABVN, nucleus tractus solitaries, and other nuclei in the brain participating cardiovascular control and eventually activating the efferent vagal fibers. Plenty of researches had proven that stimulation of cervical vagus nerve could improve LV systolic function, prevent progressive LV enlargement in dogs with heart failure, prevent sudden death in conscious dogs with a healed MI, and attenuate ventricular dysfunction and infarct size.
during acute ischemia-reperfusion injury. The present study reveals that LL-TS has the same effects as invasive cervical VNS on the vagal efferent fibers, which are part of the final pathway responsible for the regulation of intrinsic cardiac autonomic nervous system. Electric stimulation of ABVN could augment vagal tone by means of activating the afferent vagal nerve and modulate intrinsic cardiac autonomic nervous system to attain cardio-protective effect.

Clinical Implications

The beagle dog model of MI used in this study manifested many of the hemodynamic and neurohormonal phenomena in post-ischemic diseases observed in humans (i.e., marked and progressive depression of LV systolic, diastolic function and ventricular fibrosis and remodeling). Also transcutaneous electric stimulation of ABVN had already been used to treat epilepsy and depression, reduce the amount of anesthetic used during operative procedures, and suppress sepsis in a murine model of endotoxemia. The findings from this study offer an important step forward for translating the results to clinical applications. The therapeutic modality used in the present study also brought delightful results by manipulation of autonomic tone through vagal afferent nerve. We therefore propose a new neural interface approach to optimize cardiac autonomic tone for the treatment of post-ischemic heart diseases.

Study Limitations

We did not measure the neurotransmitters and associated signal transduction pathways, which may elucidate the mechanism underlying the treatment effects; further researches need to be done. The absence of background therapy with angiotensin-converting enzyme inhibitors and angiotensin II type 1 receptor antagonists in rats with heart failure. Role of kinins and angiotensin II type 2 receptors. J Clin Invest. 1997;99:1926–1935. Also transcutaneous electric stimulation of ABVN had already been used to treat epilepsy and depression, reduce the amount of anesthetic used during operative procedures, and suppress sepsis in a murine model of endotoxemia. The findings from this study offer an important step forward for translating the results to clinical applications.

Disclosures

None.

References


CLINICAL PERSPECTIVE

Therapy for cardiac dysfunction after myocardial infarction has improved over decades, but the effects of drugs remain limited and post-myocardial infarction heart failure still remains a leading cause of death and disability. Chronic vagus nerve stimulation could suppress the cardiac sympathetic outflow and prevent and reverse cardiac remodeling to improve the treatment for heart failure. However, vagus nerve stimulation treatment has to implant neurostimulator system by surgery and may cause some side effects, such as neck pain, coughing, swallowing difficulty, and voice alteration along with nausea and indigestion. In this investigation, we examined the effects of a noninvasive therapeutic modality to substitute for vagus nerve stimulation. In the beagle dog model of post-myocardial infarction cardiac dysfunction, chronic intermittent low-level transcutaneous electric stimulation of auricular branch of vagus nerve could improve cardiac function, alleviate cardiac fibrosis, and attenuate left ventricular remodeling. The findings from this study offer an important step forward for translating the results to clinical applications. Besides, transcutaneous electric stimulation of auricular branch of vagus nerve had already been used to treat epilepsy and depression in clinical practices. We therefore propose a new neural interface approach to optimize cardiac autonomic tone. These distinguishing features that underlie the potential clinical significance of this therapeutic approach provide us a novel, noninvasive approach to treat post-ischemic cardiac dysfunction.
Chronic Intermittent Low-Level Transcutaneous Electrical Stimulation of Auricular Branch of Vagus Nerve Improves Left Ventricular Remodeling in Conscious Dogs With Healed Myocardial Infarction

Zhuo Wang, Lilei Yu, Songyun Wang, Bing Huang, Kai Liao, Gaowa Saren, Tuantuan Tan and Hong Jiang

_Circ Heart Fail_. 2014;7:1014-1021; originally published online October 20, 2014; doi: 10.1161/CIRCHEARTFAILURE.114.001564

_Circulation: Heart Failure_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2014 American Heart Association, Inc. All rights reserved.
Print ISSN: 1941-3289. Online ISSN: 1941-3297

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circheartfailure.ahajournals.org/content/7/6/1014

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation: Heart Failure_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation: Heart Failure_ is online at:
http://circheartfailure.ahajournals.org//subscriptions/